

Data analysis & modelling of DNA microarray data in in genetic expression profiling

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This work is part of a project run under the auspices of two Duke Centers – the Center for Bioinformatics & Computational Biology, and the Center for Genome Technology

Web sites:

- www.isds.duke.edu
- www.isds.duke.edu/bioinformatics

(Breast Cancer) Phenotyping

Genetic features, patterns



Physiology, Clinical outcomes

Two group problems: Binary outcomes

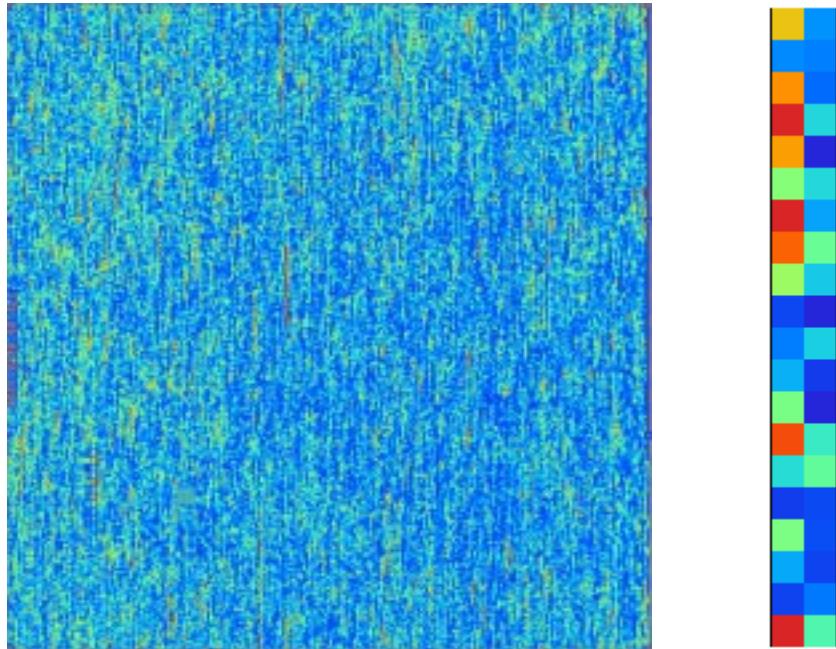
- e.g., ER+ versus ER-
- DNA microarray data: expression levels of 7129 genes (sequences) in RNA from tumour, tumour location, person, time point
- 15 ER+, 12 ER-
- Discriminatory patterns of expression?
- Predictive classification of tumours 28, 29, 30, ... ? – Decision aid
- Which genes are implicated? Surprises? What is new?

Expression array data

Microarray data: Affymetrix arrays

- 20 probe pair sets per gene: $7129 \times 20 \times 2$ intensities
- definitions of expression
- several summary measures
 - (Average) difference
 - (Average) normalised difference
 - (Average) Log Ratio – “fold” changes, level independent
- data issues: imaging, uncertain “manipulation”

One array, one probe set



Summary expression data

$p = 7126$ genes, $n = 27$ arrays

$$\mathbf{X} = [\mathbf{x}_1, \mathbf{x}_2, \dots, \mathbf{x}_n] = \begin{pmatrix} x_{1,1} & x_{1,2} & \cdots & x_{1,n} \\ x_{2,1} & x_{2,2} & \cdots & x_{2,n} \\ \vdots & \vdots & \ddots & \vdots \\ \vdots & \vdots & \ddots & \vdots \\ \vdots & \vdots & \ddots & \vdots \\ x_{p,1} & x_{p,2} & \cdots & x_{p,n} \end{pmatrix}$$

p (variables) $>>$ n (observations)

Understanding summary expression data

Ideas from latent factor modelling

e.g., on microarrays $j = 1, \dots, 15$ and $k = 16, \dots, 27$,

$$\mathbf{x}_j = \begin{pmatrix} x_{1,j} \\ x_{2,j} \\ \vdots \\ \vdots \\ \vdots \\ x_{p,j} \end{pmatrix} = \begin{pmatrix} 5 \\ \vdots \\ 5 \\ 50 \\ \vdots \\ 50 \end{pmatrix} + \begin{pmatrix} \epsilon_{1,j} \\ \epsilon_{2,j} \\ \vdots \\ \vdots \\ \vdots \\ \epsilon_{p,j} \end{pmatrix} \quad \text{and} \quad \mathbf{x}_k = \begin{pmatrix} x_{1,k} \\ x_{2,k} \\ \vdots \\ \vdots \\ \vdots \\ x_{p,k} \end{pmatrix} = \begin{pmatrix} -5 \\ \vdots \\ -5 \\ -50 \\ \vdots \\ -50 \end{pmatrix} + \begin{pmatrix} \epsilon_{1,k} \\ \epsilon_{2,k} \\ \vdots \\ \vdots \\ \vdots \\ \epsilon_{p,k} \end{pmatrix}$$

Example of factor structure (continued)

$$\mathbf{X} = \mathbf{ab}' + \mathbf{E}$$

$$\mathbf{X} = \begin{pmatrix} 5 \\ \vdots \\ \vdots \\ 50 \\ \vdots \\ \vdots \\ 50 \end{pmatrix} (1, \ 1, \ \dots, \ 1, \ -1, \ , -1, \ , \dots, \ -1) + \mathbf{E}$$

a relates subgroups of genes; **b** relates subgroups of microarrays;
E is what's left over

Factor structure in data matrices

Reality is more complicated,

$$\mathbf{X} = \mathbf{a}_1 \mathbf{b}'_1 + \mathbf{a}_2 \mathbf{b}'_2 + \cdots + \mathbf{a}_k \mathbf{b}'_k + \mathbf{E}$$

- $\mathbf{a}_1, \mathbf{a}_2, \dots$ represent patterns/relationships among genes
- $\mathbf{b}_1, \mathbf{b}_2, \dots$ represent patterns/relationships among arrays
- \mathbf{E} is what's left over ...

If \mathbf{E} is small, $\mathbf{X} \approx \mathbf{AB}$

- \mathbf{A} has columns $\mathbf{a}_1, \mathbf{a}_2, \dots$
- \mathbf{B} has rows $\mathbf{b}'_1, \mathbf{b}'_2, \dots$

Identifying decompositions: $\mathbf{B} = \mathbf{DF}$

– \mathbf{F} orthogonal, \mathbf{A} orthonormal columns, \mathbf{D} diagonal –

Singular value (factor) decomposition

$$\mathbf{X} = \mathbf{ADF}$$

$$\begin{pmatrix}
 x_{1,1} & \cdots & x_{1,n} \\
 x_{2,1} & \cdots & x_{2,n} \\
 \vdots & \ddots & \vdots \\
 x_{p,1} & \cdots & x_{p,n}
 \end{pmatrix} =
 \begin{pmatrix}
 a_{1,1} & \cdots & a_{1,n} \\
 a_{2,1} & \cdots & a_{2,n} \\
 \vdots & \ddots & \vdots \\
 a_{p,1} & \cdots & a_{p,n}
 \end{pmatrix} \begin{pmatrix}
 d_1 & 0 & \cdots & 0 \\
 0 & d_2 & \cdots & 0 \\
 \vdots & \vdots & \ddots & 0 \\
 0 & 0 & \cdots & d_n
 \end{pmatrix} \begin{pmatrix}
 f_{1,1} & \cdots & f_{1,n} \\
 f_{2,1} & \cdots & f_{2,n} \\
 \vdots & \ddots & \vdots \\
 f_{n,1} & \cdots & f_{n,n}
 \end{pmatrix}$$

Factor decompositions (continued)

$$\mathbf{X} = \mathbf{ADF}$$

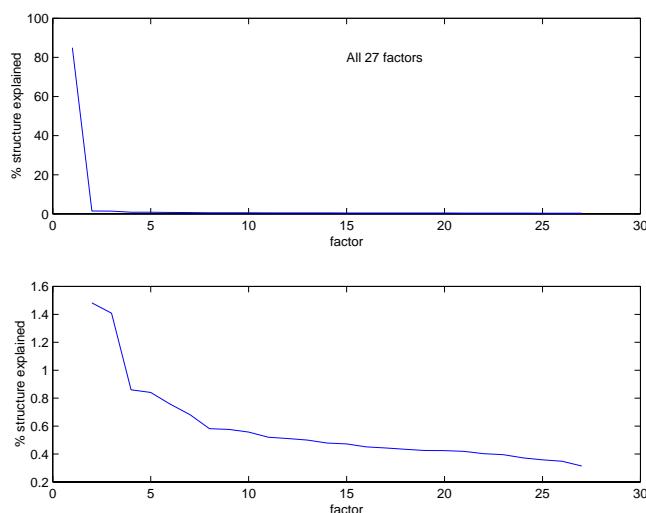
- **Factor loadings matrix \mathbf{A}** has columns $\mathbf{a}_1, \mathbf{a}_2, \dots$
 - patterns/relationships among genes
- **Latent factors** are rows of \mathbf{F}
 - patterns/relationships among arrays
- **Relative importance** of factors 1, 2, are $d_j > 0$
 - large d_1, \dots small d_n
- **Supergenes**=Factors are linear combinations of expression
 - microarray j : column j of \mathbf{F} is $\mathbf{f}_j = \mathbf{D}^{-1} \mathbf{A}' \mathbf{x}_j$

Principal components analysis: eigenvalues/vectors of (singular) \mathbf{XX}'

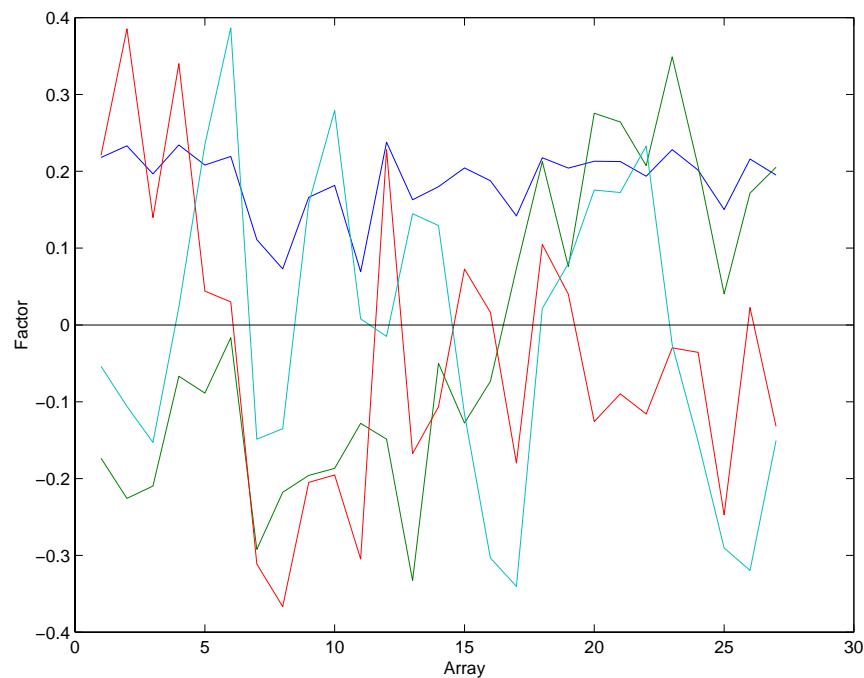
SVD of breast cancer data matrix

\mathbf{X} is 7129×27 , \mathbf{A} is 7129×27 , \mathbf{D} is diagonal 27×27 , \mathbf{F} is 27×27

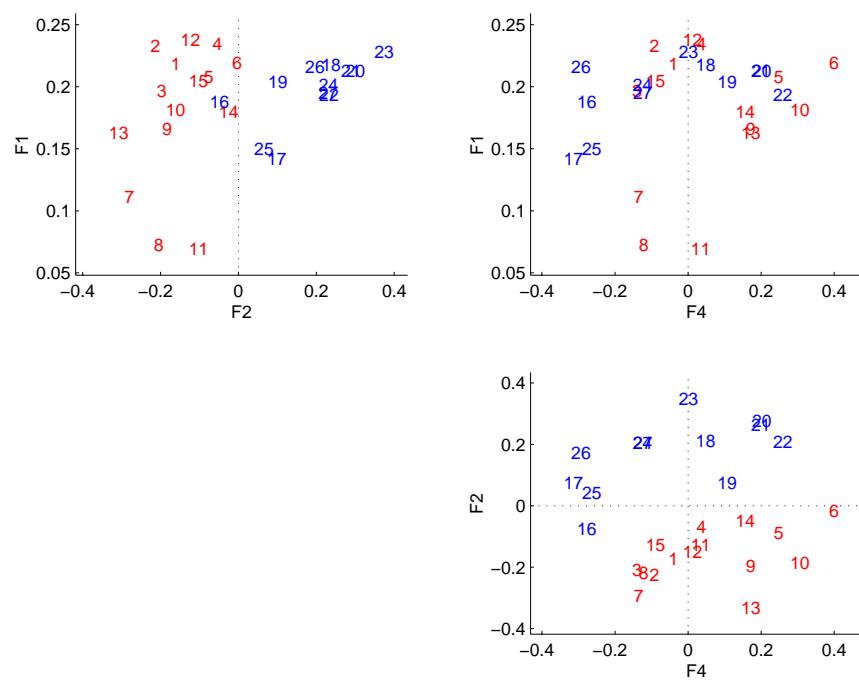
Relative contributions of factors – d_j^2 as % of total:



4 of the factors across arrays



Arrays plotted on pairs of 3 factors



Binary regression modelling

- Microarray j , expression profile \mathbf{x}_j
- Binary classification: 1 (ER+) or 0 (ER-)
- Probability array j is ER+ is $p(\mathbf{x}_j)$
- Standard “probit” model: $p(\mathbf{x}_j) = \Phi(\mathbf{x}'_j \boldsymbol{\beta})$
- Linear regression on gene expression, mapped to probability scale
 - $\mathbf{x}'_j \boldsymbol{\beta} = \sum_{i=1}^p \beta_i x_{i,j}$
 - β_i is regression coefficient on gene i

Binary regression modelling: $p >> n$

$n = 27$ binary observations, $p = 7129$ primary covariates

Data reduction? **Sufficient** data reduction?

$$[\mathbf{x}'_1 \boldsymbol{\beta}, \mathbf{x}'_2 \boldsymbol{\beta}, \dots, \mathbf{x}'_n \boldsymbol{\beta}] = \mathbf{X}' \boldsymbol{\beta}$$

or

$$(\mathbf{D}\mathbf{F})' \boldsymbol{\gamma}$$

with

$$\boldsymbol{\gamma} = \mathbf{A}' \boldsymbol{\beta}$$

Regression on genes (in \mathbf{X}) translates to regression on **supergenes** – the factor variables in \mathbf{F} – weighted by elements of \mathbf{D}

Regression on supergenes: Priors

- Supergene (factor) model has $n = 27$ orthogonal covariates
- Precisions d_j^2 related to data on element γ_j of $\boldsymbol{\gamma}$
- Suitable priors on $\boldsymbol{\gamma}$
 - elements γ_j independent (orthogonality)
 - conditionally conjugate: $\gamma_j \sim N(0, \tau_j^2/d_j^2)$
 - differing scales τ_j
 - hyperprior on τ_j

Prior on $\boldsymbol{\gamma} = \mathbf{A}\boldsymbol{\beta}$ must be derived from prior on $\boldsymbol{\beta}$

Regression on genes: Priors

Prior: $p(\boldsymbol{\beta})$

- Generalised singular “g-prior”
- “Shape” defined by \mathbf{A} , “scales” to be estimated

$$p(\boldsymbol{\beta}) \propto \exp\{-\boldsymbol{\beta}'(\mathbf{A}\mathbf{T}^{-1}\mathbf{D}\mathbf{A}')\boldsymbol{\beta}/2\}$$

with

$$\mathbf{T} = \text{diag}(\tau_1^2, \dots, \tau_n^2)$$

Standard g-prior: $\tau_j^2 = \text{constant}$, and prior non-singular

Regression on genes via supergenes

Prior for β : centred at $\mathbf{0}$

- neutral: implied classification probability of 0.5
- multiple solutions to $\gamma = \mathbf{A}'\beta$
- ω in null space of \mathbf{A} : $\mathbf{A}'\beta = \mathbf{A}'(\beta + \omega)$
- same posteriors for priors on β with mean ω
- choose $\omega = \mathbf{0}$ to identify unique prior, unique posterior

Posterior for β : Defined uniquely by $\beta = \mathbf{A}\gamma$

- 27 supergene parameters \rightarrow 7129 gene parameters
- Posterior simulation samples for β
- Posterior mean, significance of genes, effects of genes

Latent normals in probit regression

$$p(\mathbf{x}_j) = Pr(\text{array } i \text{ is } ER+) = Pr(y_i \geq 0)$$

where

$$y_i \sim N(\mathbf{x}'_i \beta, 1)$$

or, as vector for all arrays,

$$\mathbf{y} \sim N(\mathbf{X}'\beta, \mathbf{I})$$

or

$$\mathbf{y} \sim N(\mathbf{F}'\mathbf{D}\gamma, \mathbf{I})$$

- critical to model fitting/computation
- interpretation and implied *kernel regression structure*

Kernel regression structure under GSG prior

Marginalising over β implies

$$\mathbf{y} \sim N(\mathbf{0}, \mathbf{K})$$

with *kernel covariance matrix*

$$\mathbf{K} = \mathbf{F}'\mathbf{T}\mathbf{F} + \mathbf{I}$$

with

$$\mathbf{T} = \text{diag}(\tau_1^2, \dots, \tau_n^2)$$

- correlations between arrays
- effective dependence structure with respect to classification
- key role of \mathbf{T}
- effective *non-linear classifier*

Regression analysis & computation

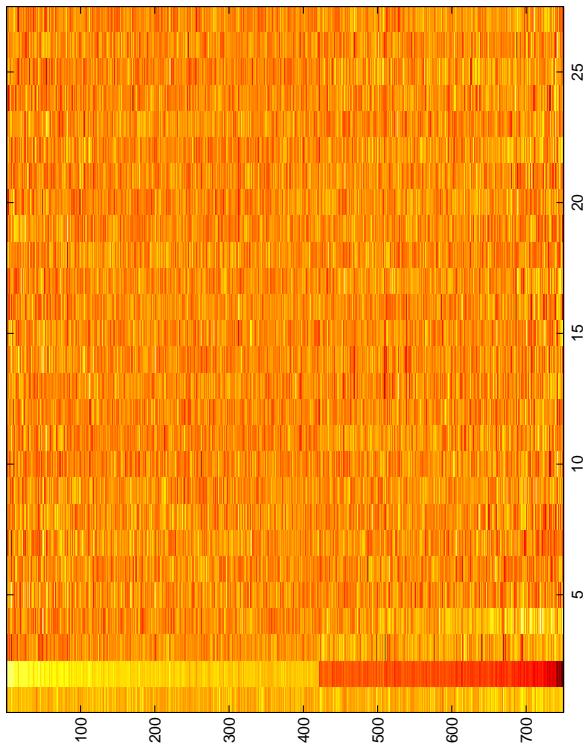
Prior: $\prod_{j=1}^n p(\gamma_j | \tau_j) p(\tau_j)$

- Posterior: **Easy** MCMC: iterative simulation to impute
 - latent normals \mathbf{y} underlying probit
 - each γ_j, τ_j
- Hierarchical: prior on τ_j^2 – sensitivity to hyperparameters
- Variants: model selection priors, extra-probit variation, ...

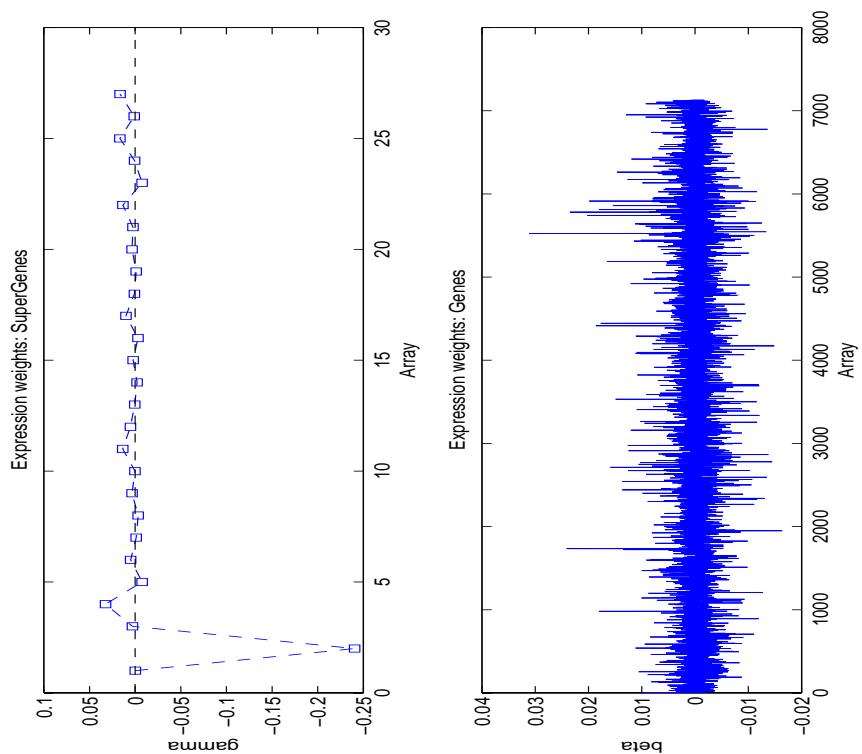
Out-of-sample or **Cross-validatory** prediction:

- New tumour on microarray – data \mathbf{x} : Posterior for $p(\mathbf{x}) = \Phi(\mathbf{x}'\beta)$
- One-at-a-time analysis: Treat array i as having “Missing” binary indicator

Factor loadings A for top 750 genes



Estimated regression coefficients



Top 10 genes with + beta coefficients

High in ER+, Low in ER-

... all almost surely > 0 ...

- mRNA for oestrogen receptor
- ps2 protein gene
- intestinal trefoil factor mRNA, complete cds
- nat1 gene for arylamine N-acetyltransferase
- mRNA for cardiac gap junction protein
- complement component C4a gene
- **breast cancer, estrogen regulated liv-1 protein (liv-1) mRNA**
- 5t4 gene for 5t4 oncofetal antigen
- nedd-4-like ubiquitin-protein ligase wwp1 mRNA
- hepatocyte nuclear factor-3 alpha (hnf-3 alpha) mRNA

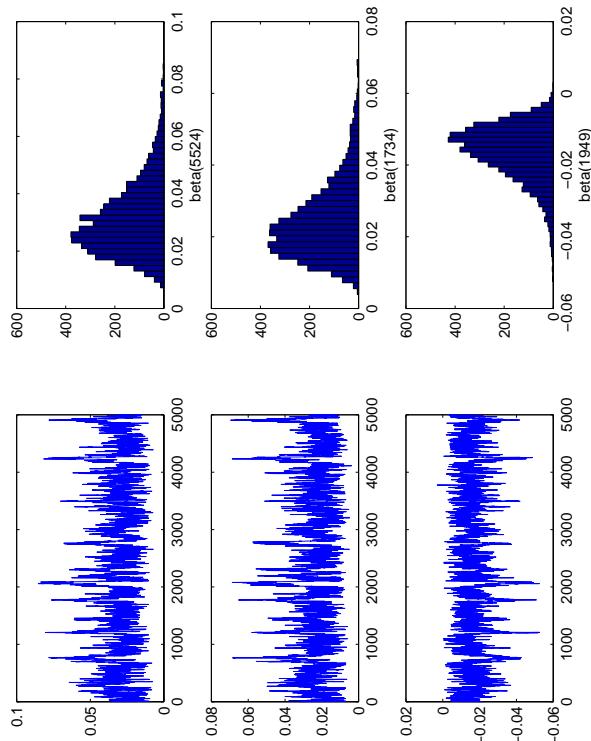
Top 8 genes with - beta coefficients

Low in ER+, High in ER-

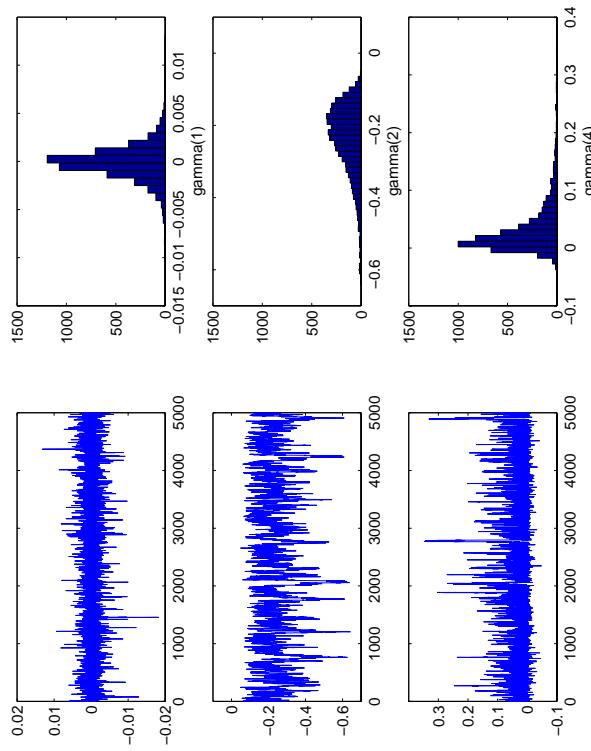
... all almost surely < 0 ...

- (hybridoma h210) anti-hepatitis A IgG variable region
- rearranged immunoglobulin lambda light chain mRNA
- Ig alpha 2=immunoglobulin A heavy chain allotype 2
- omega light chain protein 14.1 (Ig lambda chain related) gene
- sry-related HMG-box 12 protein
- matrilysin gene
- guanylate binding protein isoform I (gbp-2) mRNA
- cystic fibrosis antigen mRNA, complete cds

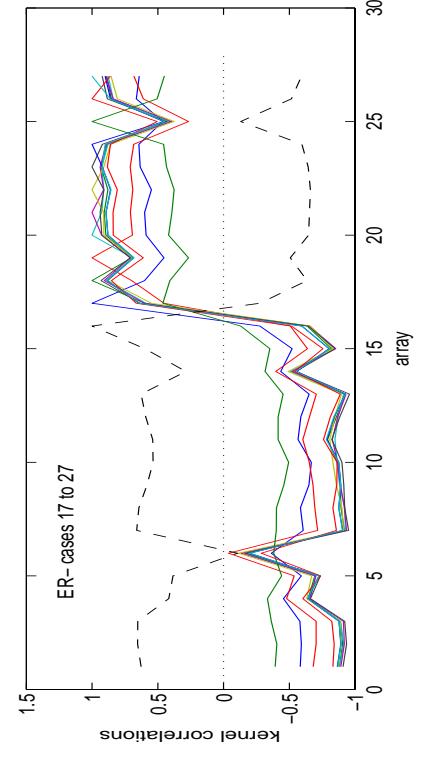
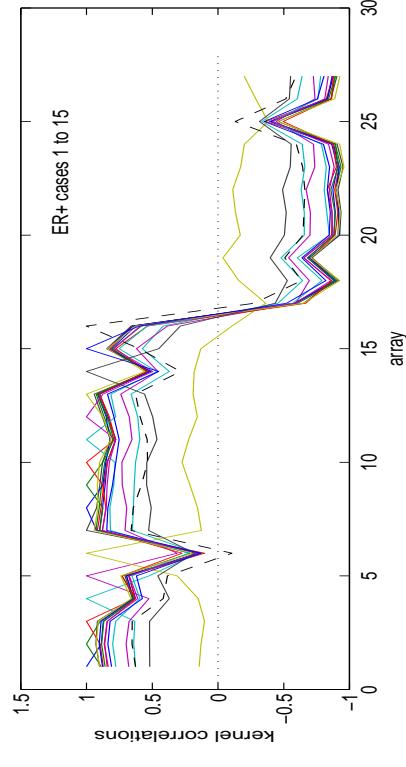
Posterior for 3 beta coefficients



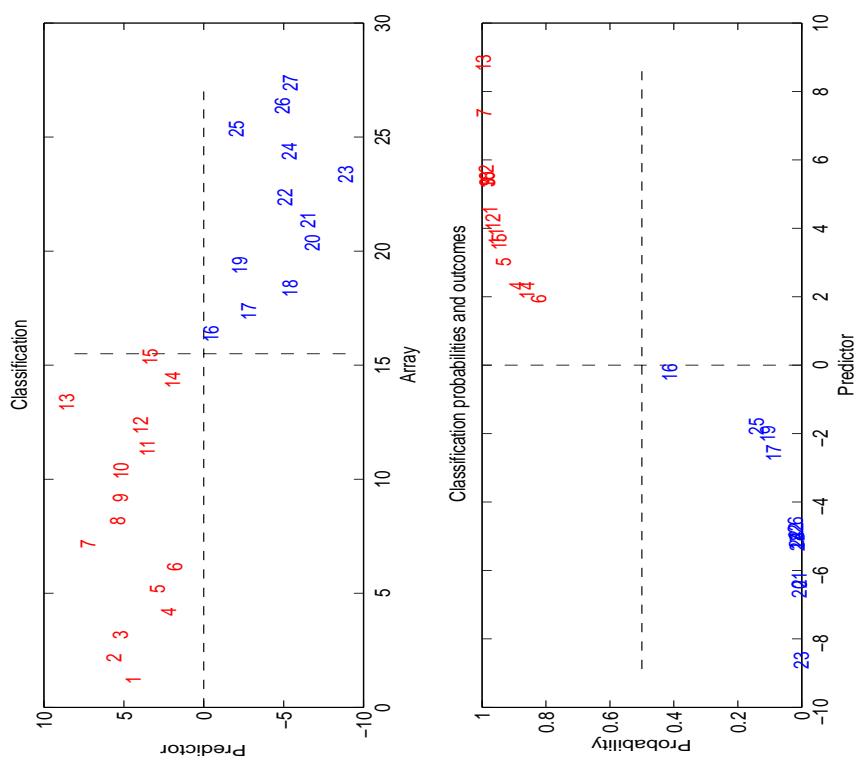
Posteriors for 3 gamma coefficients



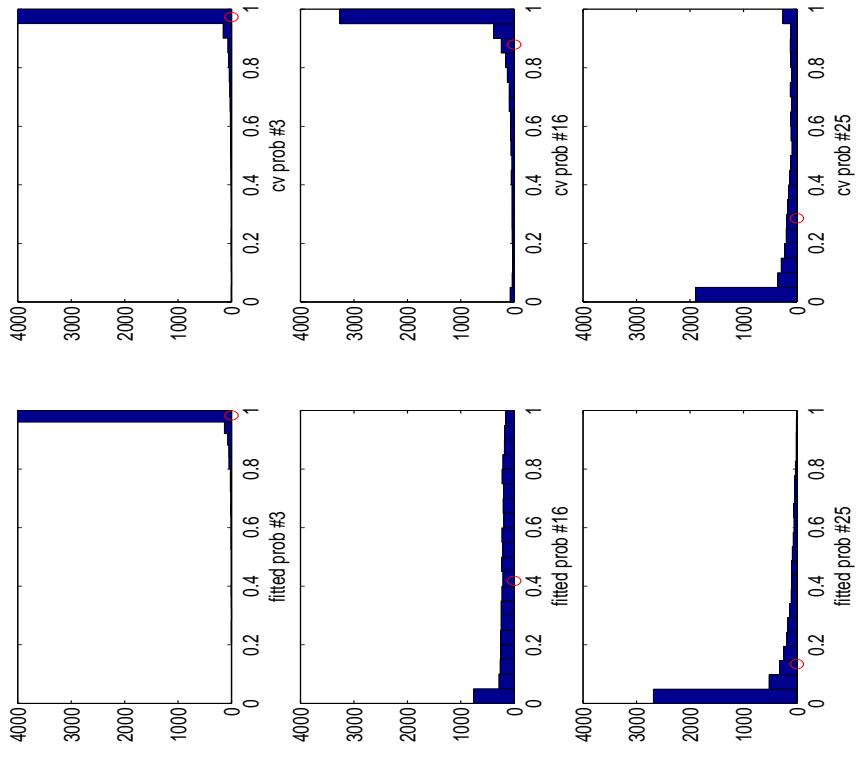
Estimated kernel correlation structure



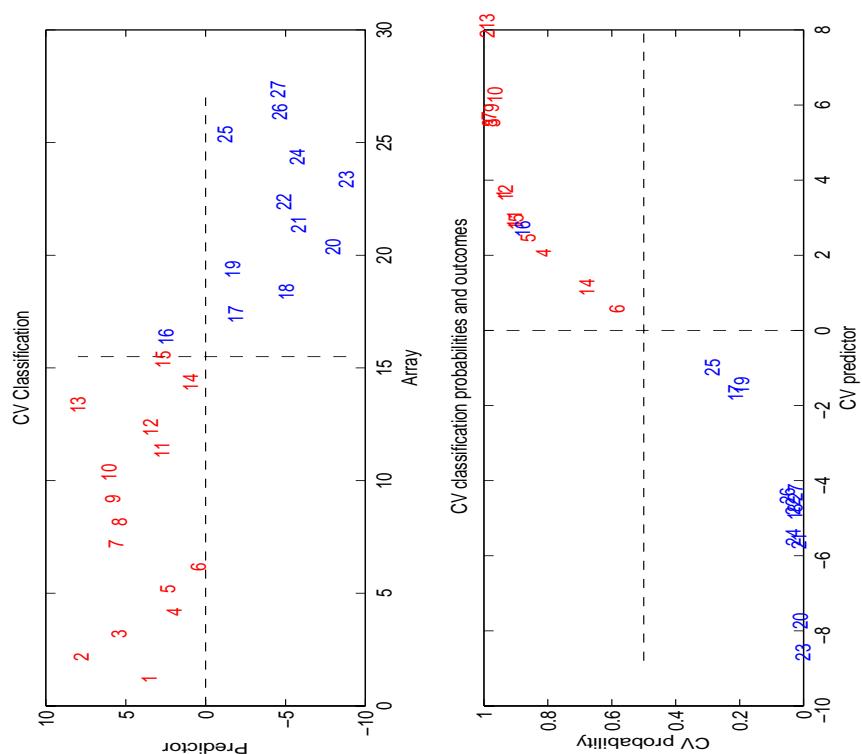
Fitted classification



Uncertainty in classification

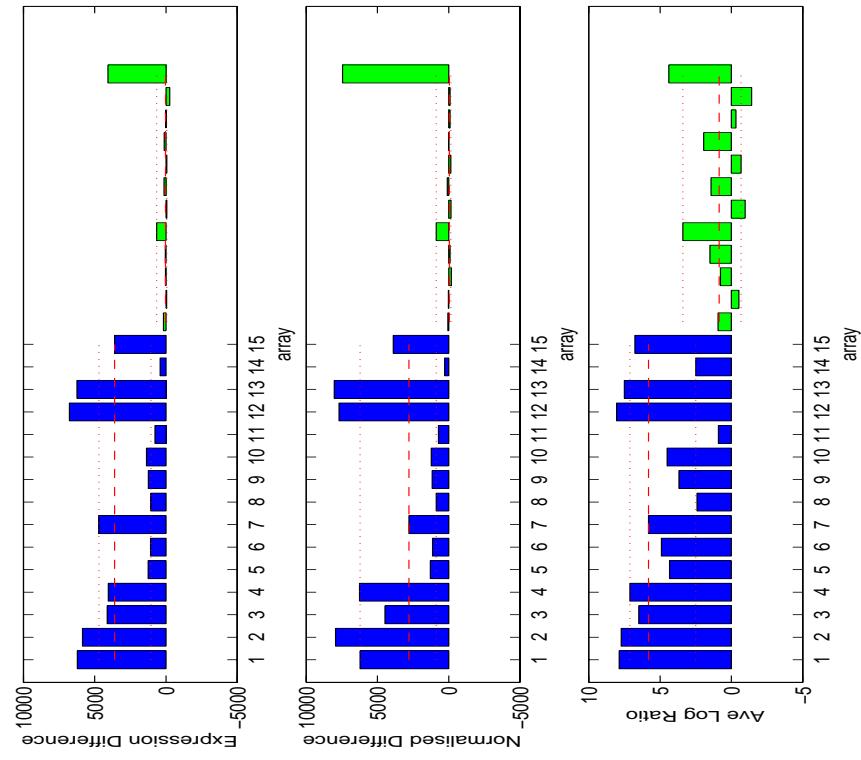


Cross-Validatory predictions

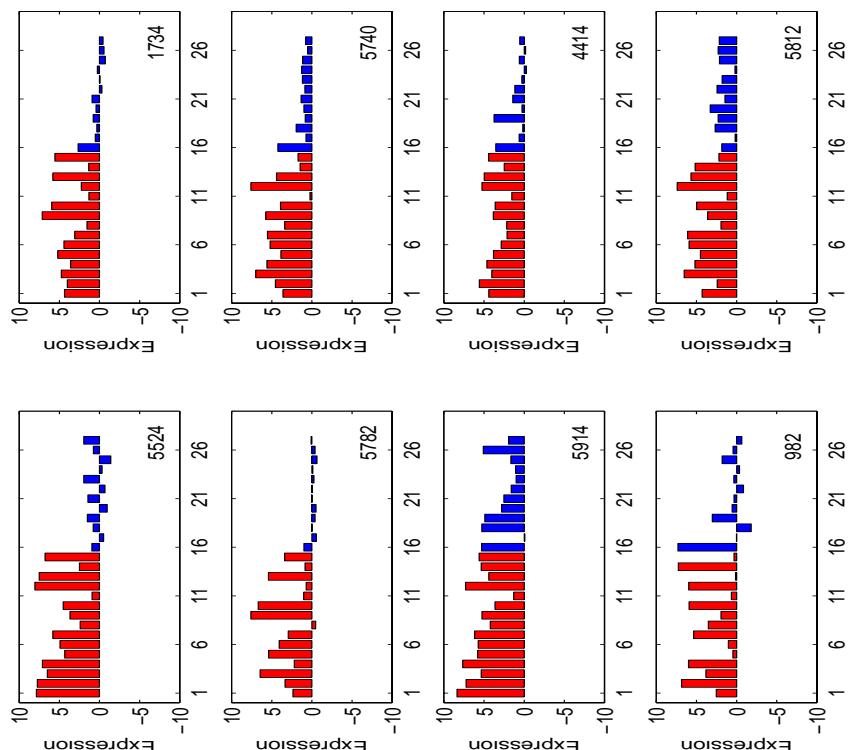


Expression measures: Top + gene

Expression Difference, Normalised difference &
Log-Ratio for oestrogen receptor gene

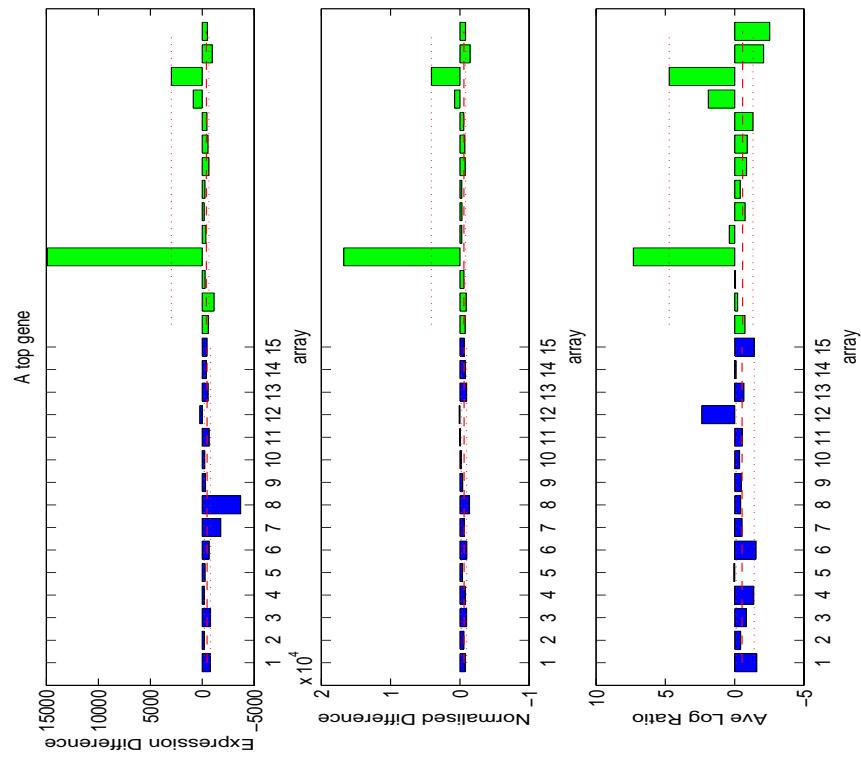


Expression by arrays: top 8(+) genes



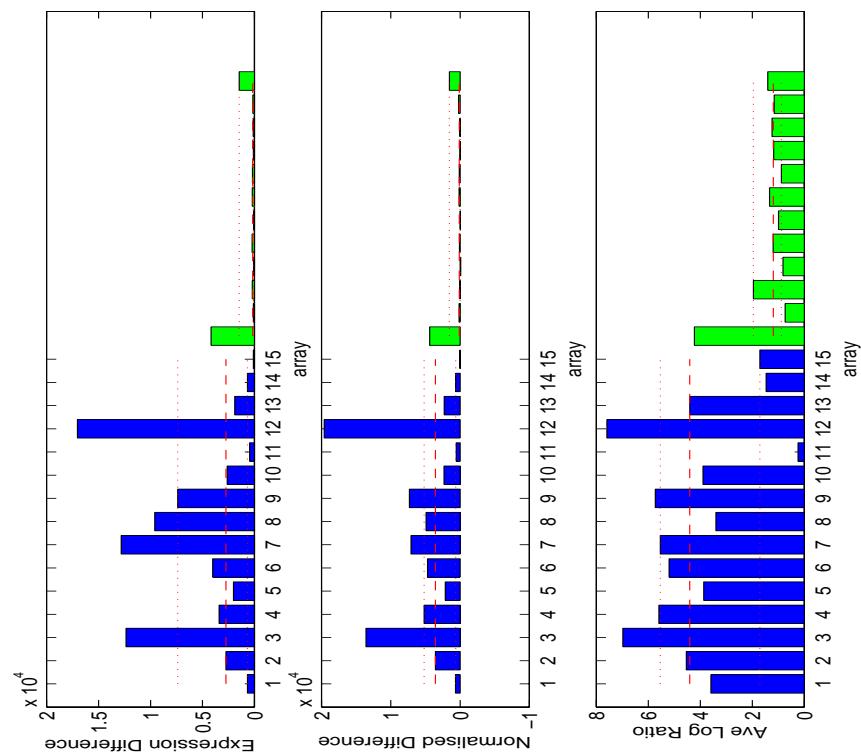
Expression measures: A - gene

Expression Difference, Normalised difference &
Log-Ratio



Expression measures: A + gene

Expression Difference, Normalised difference &
Log-Ratio



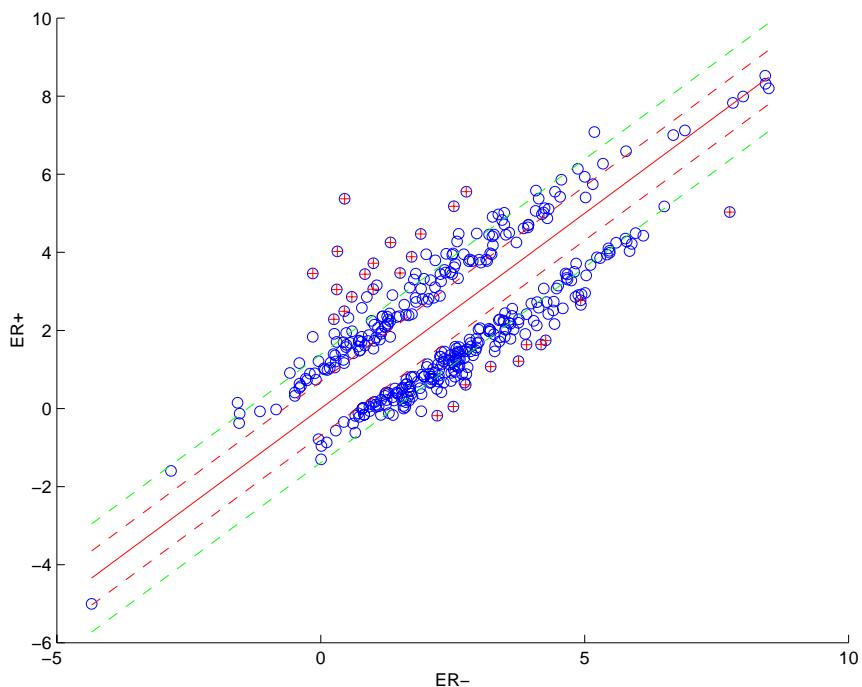
Screening for “Top” genes

- Screen genes by ordering $|E(\beta_i|Data)|$
- Select “top k ”
 - Screen and refit model –
- Summary based on top $k = 400$ genes
- Refines classification: array 16 is “better” classified
 - “top genes” favour correct classification ER+
 - some/many genes screened favour ER+
 - learning opportunities vs. simple classification
- Warns against simple screening and use of subsets of “discriminatory” genes
 - Model analysis as “filter”

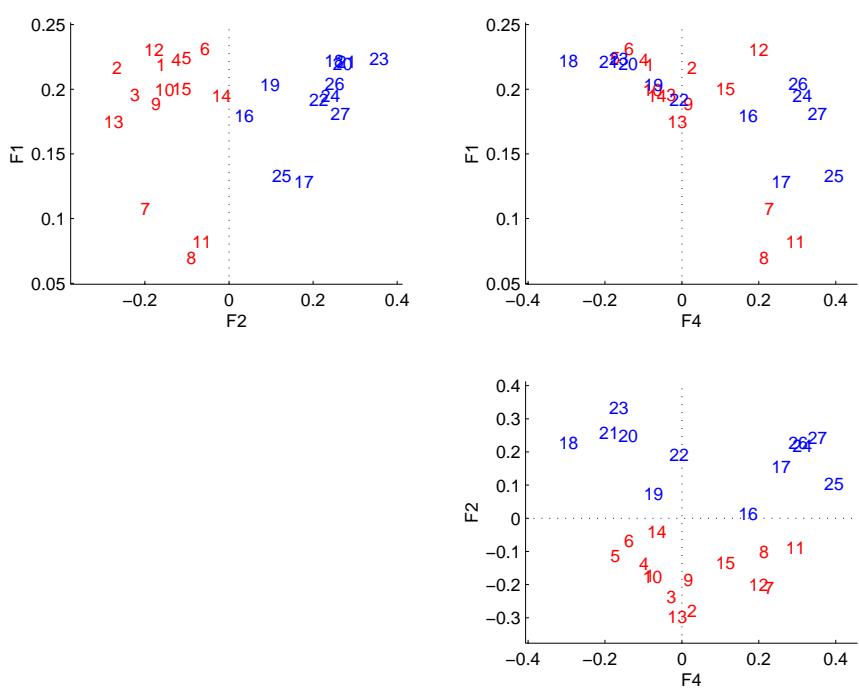
Sample 16

- ER+ or ER-?
- High uncertainty about $Pr(ER+)$
 - small sample size –
- Explore “top genes”
- Oestrogen gene appropriately “down”
- Others are “up”
 - PS2 protein gene: 2nd top gene
 - Liv-1 protein gene: 7th top gene
- Both regulated by oestrogen receptor
 - High on array 16, as are others
 - – Other regulators of PS2, Liv-1 ... ?
 - – ER status determination ... ?

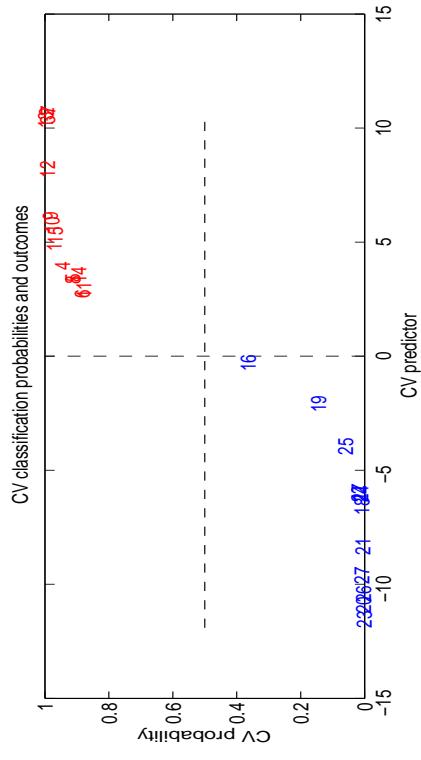
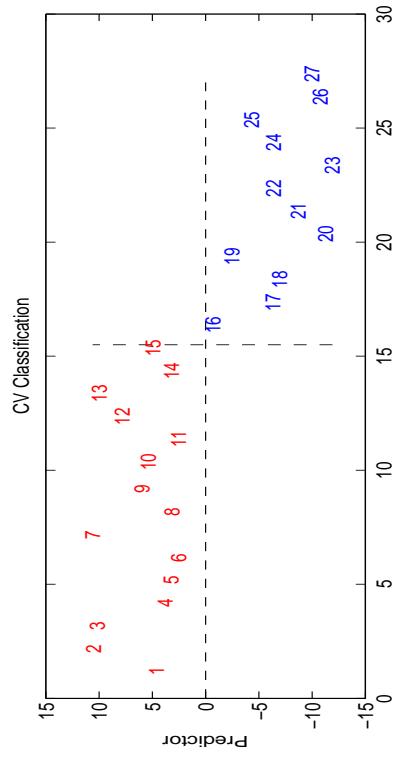
Top 400 genes: Ave(expression) on ER+ versus ER-



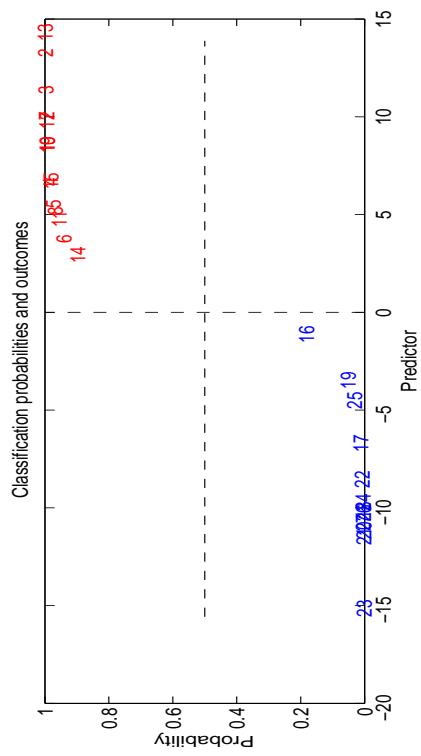
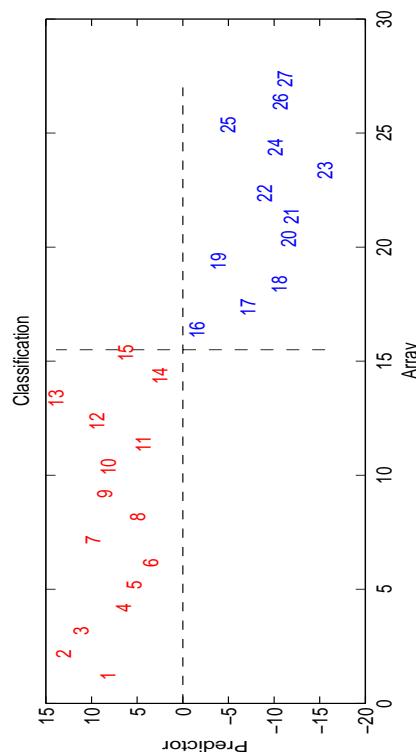
Arrays on pairs of 3 factors: Top 400 genes



CV predictions: Top 400 genes



Fitted classification: Top 400 genes



Current and Future

Application(s)

- Other binary outcomes (remission/not, ...)
- More data
- Multiple outcomes: cancer stages/states
- Multiple expression measures
- Other biomedical contexts

Methodology

- Small samples/low information:
 - MCMC convergence, prior specifications –
- Errors in variables: uncertain expression
- Multiple outcome categories: multinomials
- Non-linear regressions (kernel methods)

Uncertainty: Top 400 genes

