

Computational Pathology: Towards Precision Medicine

Andrew Janowczyk, PhD

Assistant Research Professor



CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS

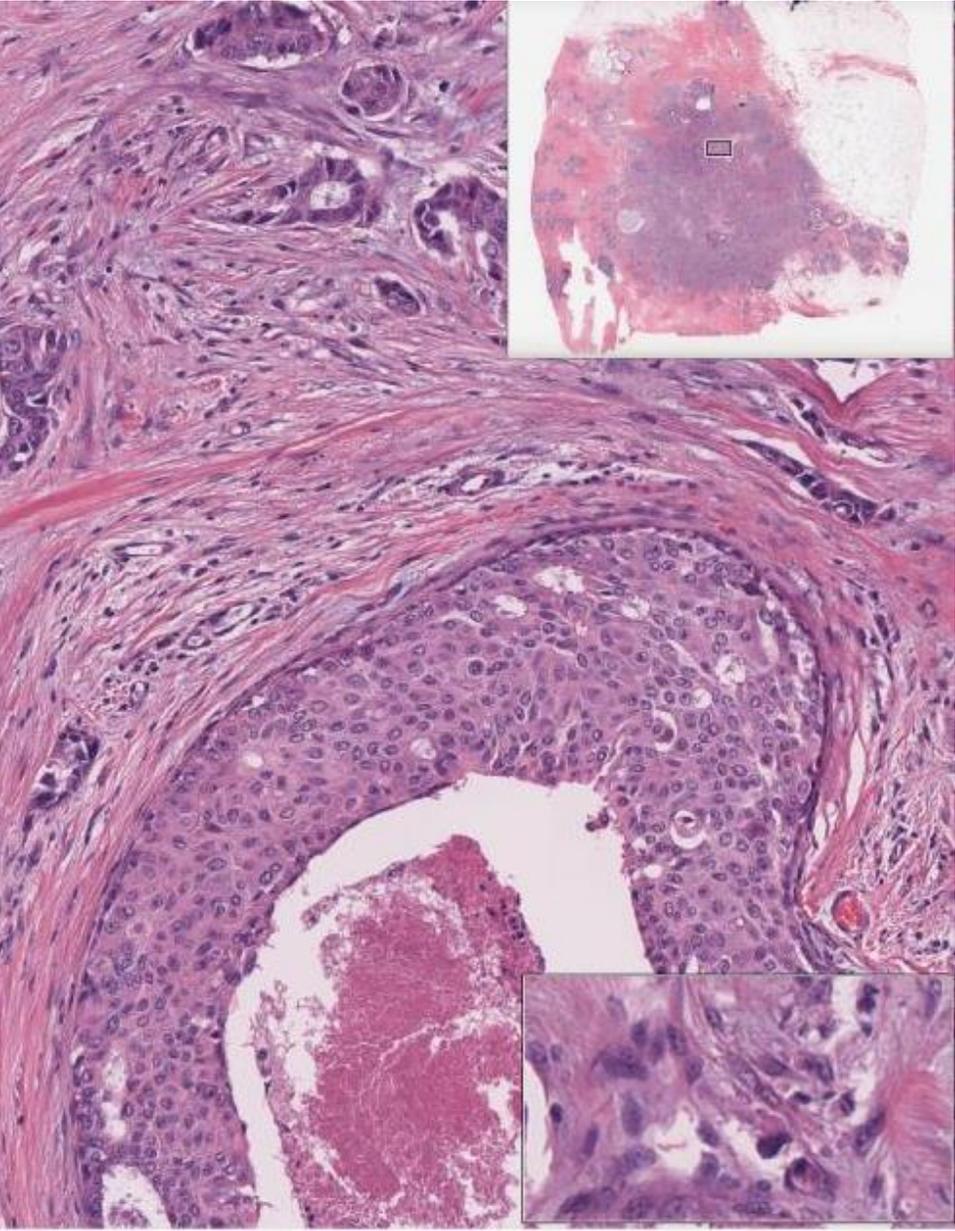


**Swiss Consortium
for Digital Pathology**

Outline

1. Introduction to Computational Digital Pathology
2. Research Applications
3. HistoSuite Tools

Brief Introduction



What is Computer Aided Diagnostics (CAD)?

- Using algorithms to help clinicians analyze data
- (f)MRI, Histology, Xray, CT, etc

Why is it useful?

- Improves efficiency & robustness of medical diagnoses
 - Fast, reproducible
- Leverage vast amounts of data already in existence
 - More being created daily at an increasing rate

How can we use this data?

- Can perform data mining to identify trends
- Identifying subtle image patterns that may not be visually discernible
- Build systems to *aid*, not replace, doctors through *decision support*
- Long term goal of leading to precision medicine

Research vs Clinical CAD Applications

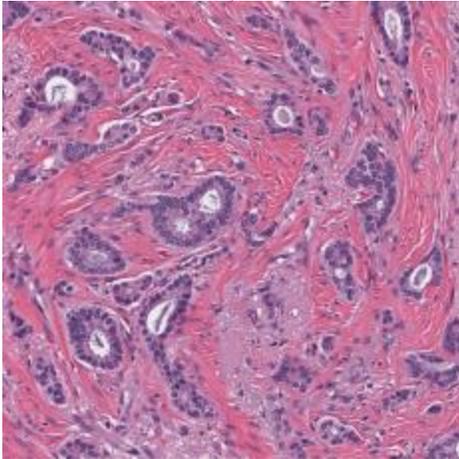
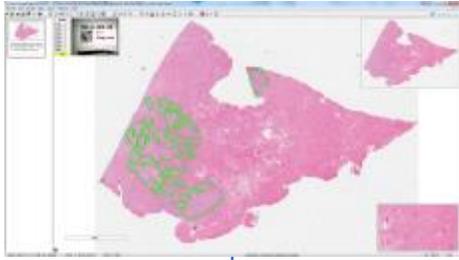
- Clinical Applications

- Recapitulate and automate existing processes
- Cancer detection, grading, counting and area estimation tasks
- Improvements through quantification, reproducibility, and definition refinement



- Research Applications

- Develop novel features and metrics
- Sub-type discovery
- Biology elucidation
- Improvements through augmentation and new insights



Nuclei
Segmentation

Stroma
Segmentation

Lymphocyte
Segmentation

Mitosis
Detection

Tubule
Segmentation

ROI
Identification

Morphology
Density
Texture

Structure
Texture
Organization

Infiltration
Status

Number per
ROI

Regularity

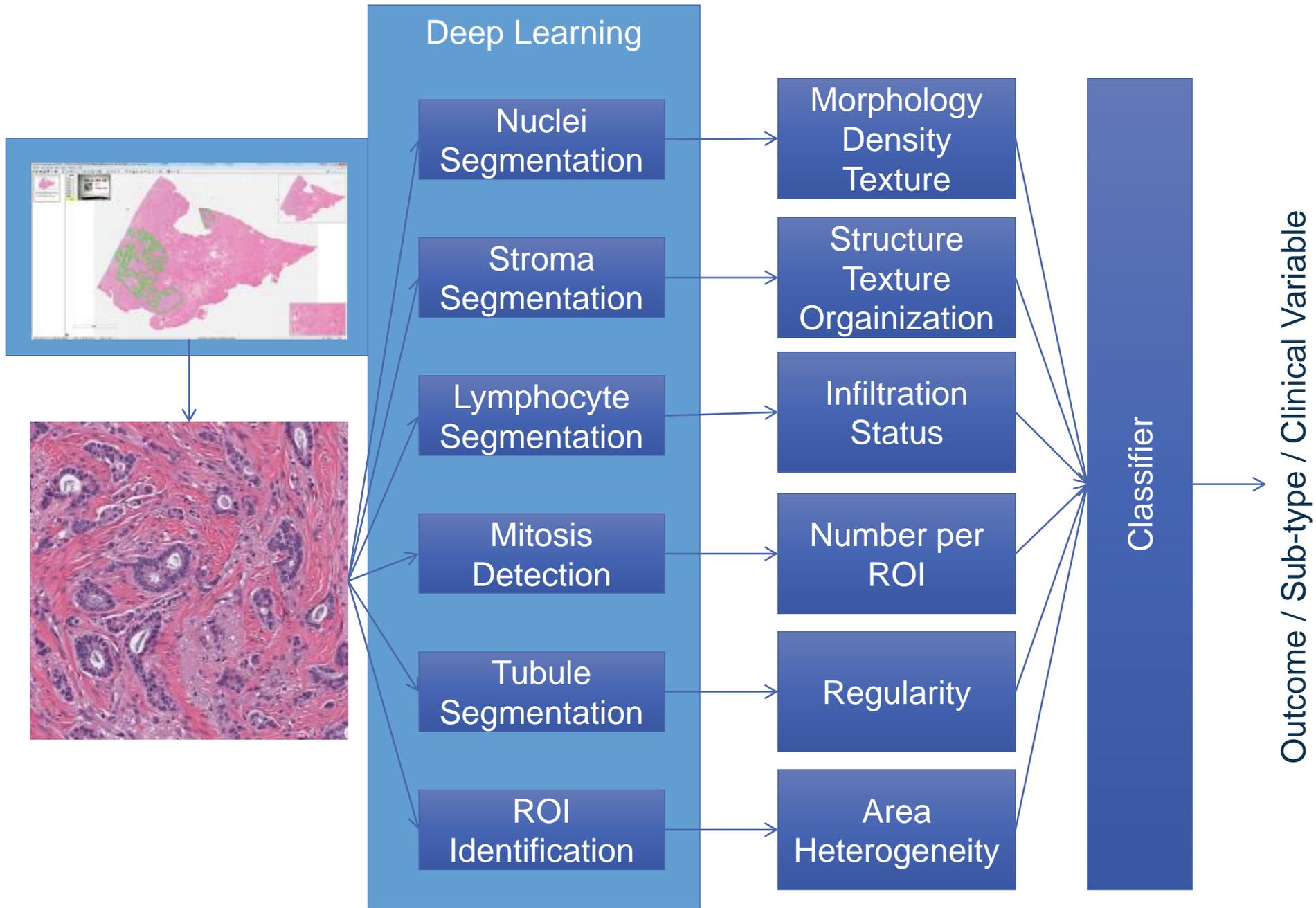
Area
Heterogeneity

Classifier

Outcome / Sub-type / Clinical Variable

Present – Deep Learning





Why use deep learning?

- Faster than creating hand-crafted features
 - Hand-crafted nuclei segmentation = 3 years
 - With deep learning = 3 hours
- Shows great robustness (both presentation and noise)
 - Able to examine more cases than developers

- At the end of the day:
 - Clinicians don't care *how the pieces were made*
 - Only care *what they can do with them*

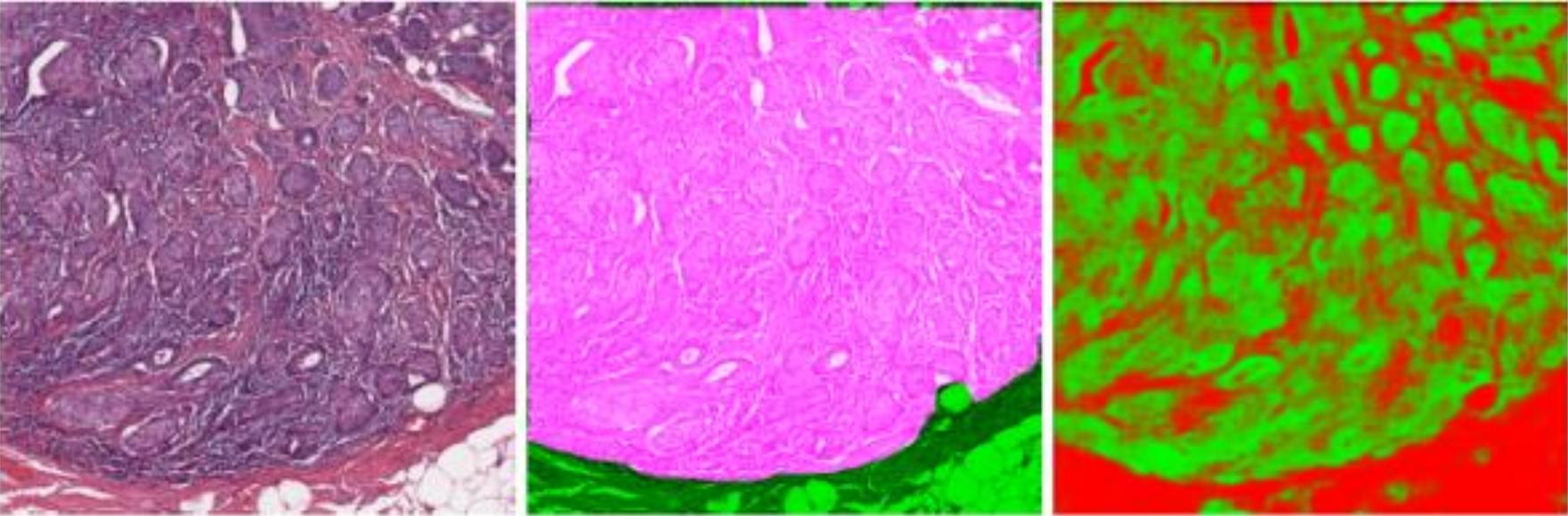
Our Proposed Framework

- “Deep learning for image analysis tasks in digital pathology: A comprehensive tutorial with selected applications in lymphoma, colorectal and breast cancer analysis”, **Andrew Janowczyk** and Anant Madabhushi, JPI 2016 (Most viewed award, >51k downloads)
- Through 7 use cases, provide best practices, code + data + tutorial for:
 1. nuclei segmentation (f-score of .83 across 12,000 nuclei)
 2. epithelium segmentation (f-score of .84 across 1,735 regions)
 3. tubule segmentation (f-score .83 from 795 tubules),
 4. lymphocyte detection (f-score .90 across 3,064 lymphocytes),
 5. mitosis detection (f-score .53 across 550 mitotic events),
 6. invasive ductal carcinoma detection (f-score .7648 on 50k testing patches)
 7. lymphoma sub-type classification (classification accuracy of .97 across 374 images)

•All results are either comparable or superior to current state of the art

Segmenting Epithelium

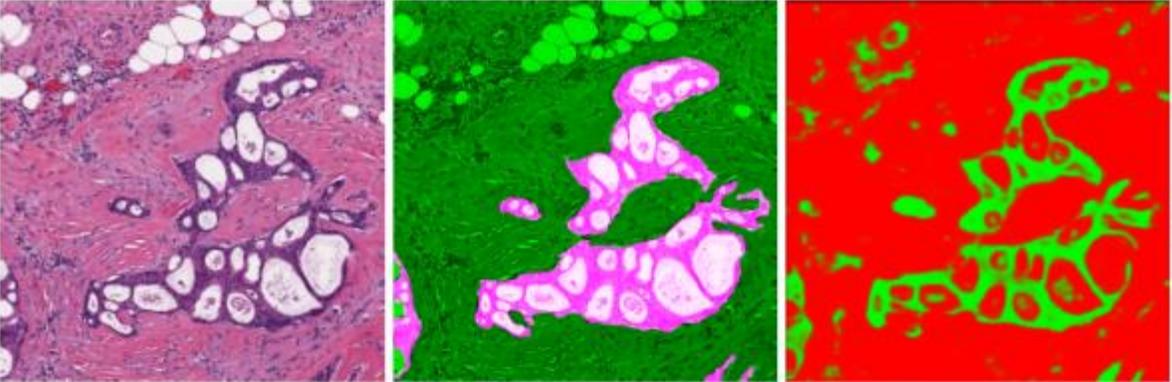
Original images in (a) and (d) with their associated ground truth in (b) and (e) overlaid in fuchsia. We can see that the results from the deep learning, in (c) and (f), that a pixel level metric is perhaps not ultimately suited to quantify this task as DL is better able to provide a pixel level classification, intractable for a human expert to parallel.



(a)

(b)

(c)



(d)

(e)

(f)

Development and evaluation of deep learning-based segmentation of histologic structures in the kidney cortex with multiple histologic stains

Study Aims:

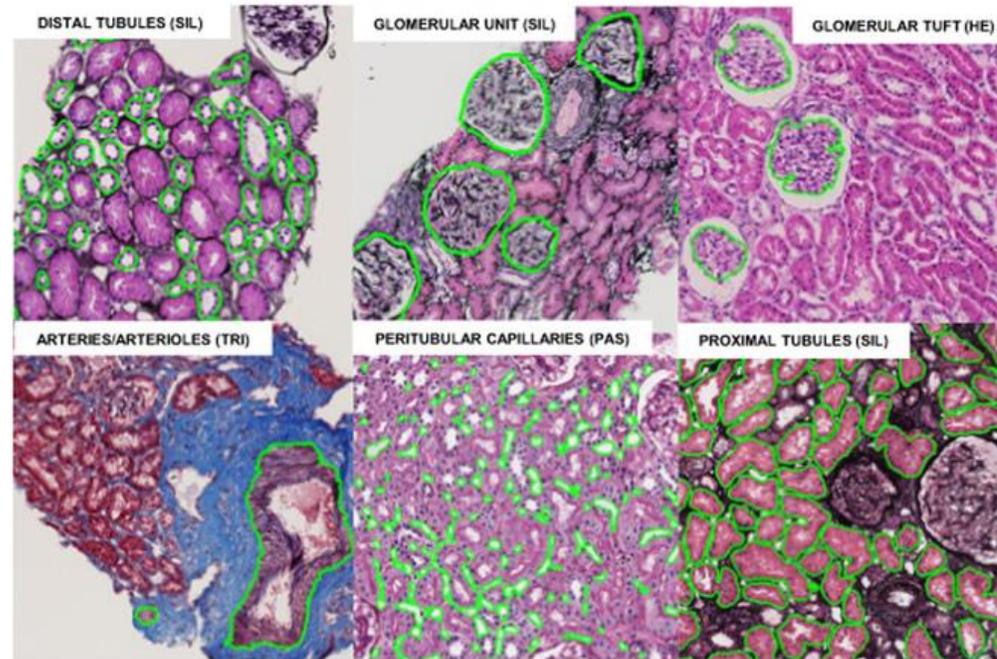
Novel protocols for renal biopsy assessment

Feasibility of deep learning-based (DL) convolutional neural networks (CNNs) for normal histology, to facilitate quantitation of prognostic histologic structures

Dataset:

- 125 NEPTUNE MCD biopsies
- H&E, PAS, TRI, SIL stains
- 459 WSIs of normal renal parenchyma (MCD)
- 38 pathology laboratories
- 30048 annotations generated across primitives

U-Net DL Segmentation using multistained WSIs



Results:

- Comparative DL performance across 4 stains (Best results on PAS stained WSIs)
- Multiple DL networks with suggested number of training exemplars across primitives
- Optimal digital magnification: 5X glomeruli, 10X tubules and arteries, 40X peritubular capillaries
- Validated on nephrectomies
- Online access to data and tutorials to setup DL networks

CONCLUSION:

DL-based CNNs permit efficient segmentation of kidney histologic structures on multiple stains with substantial tissue heterogeneity across centers. This work creates a technical foundation to support pathology workflows for better disease characterization and risk assessment.

Jayapandian, Chen et al, 2020

2. Research Applications

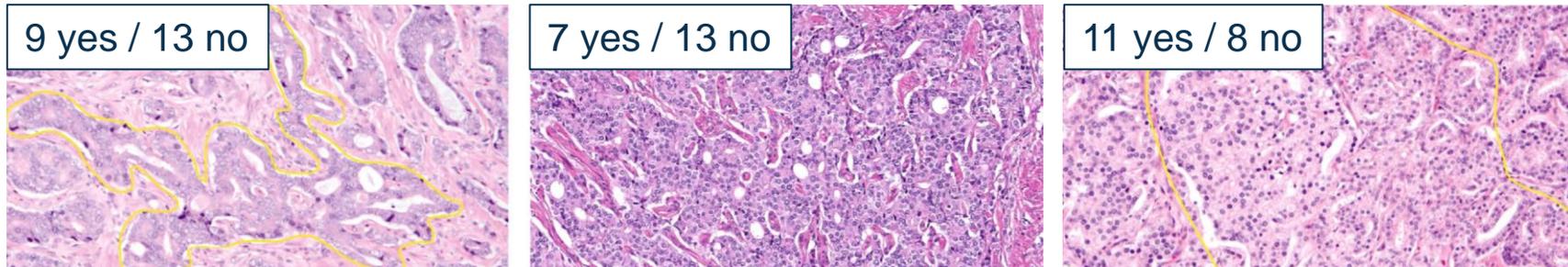


CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS

Automated cribriform quantification is prognostic of biochemical recurrence

- Cribriform increasingly recognized as high risk in prostate cancer
- Recent grading updates moved all cribriform patterns to Gleason 4
- **Computers can take the subjectivity out of cribriform assessment**

Kweldam et al. (2016) asked pathologists: Is this cribriform? Found just 23% agreement



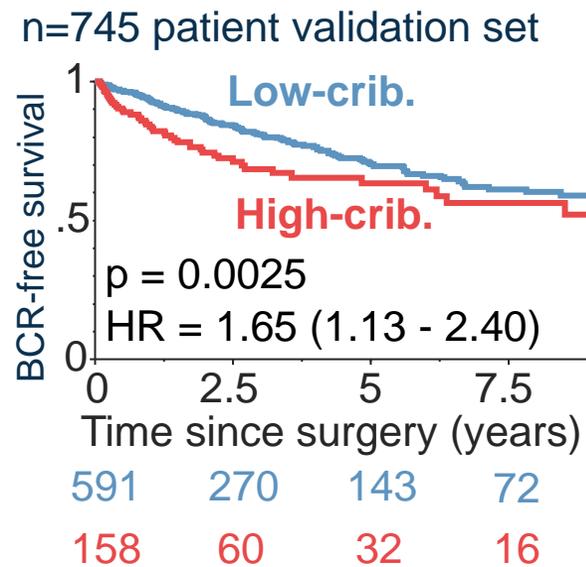
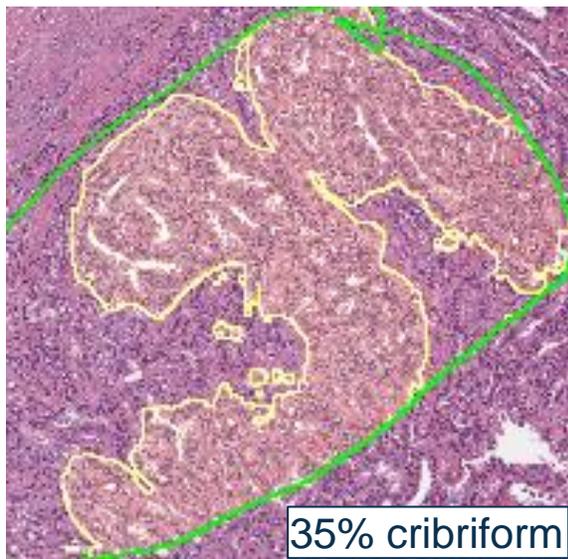
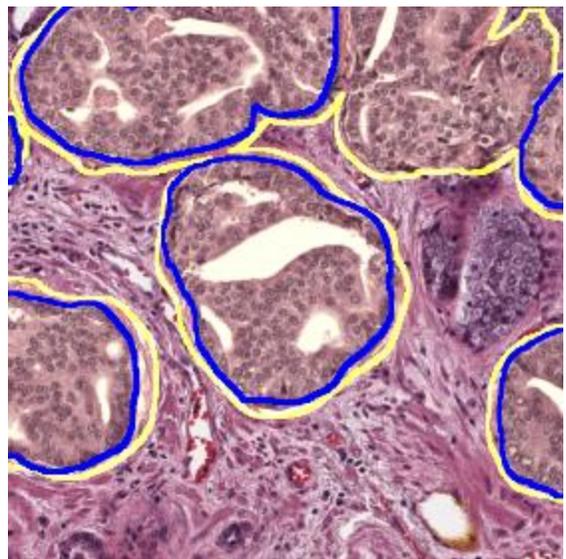
Pathologist annotations
train **deep learning model**



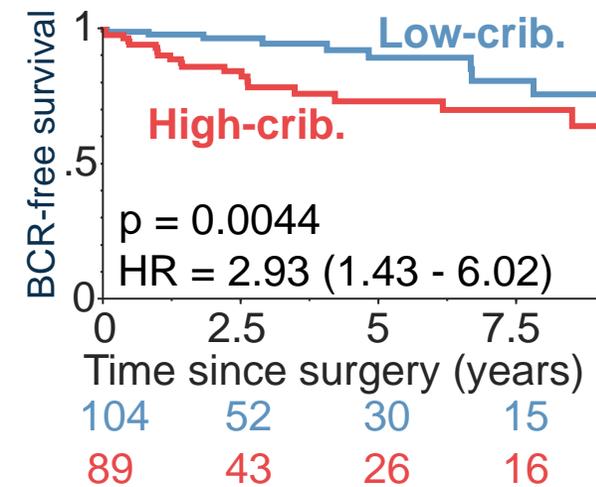
Model measures tumor
cribriform fraction



Automated cribriform assessment was prognostic in:



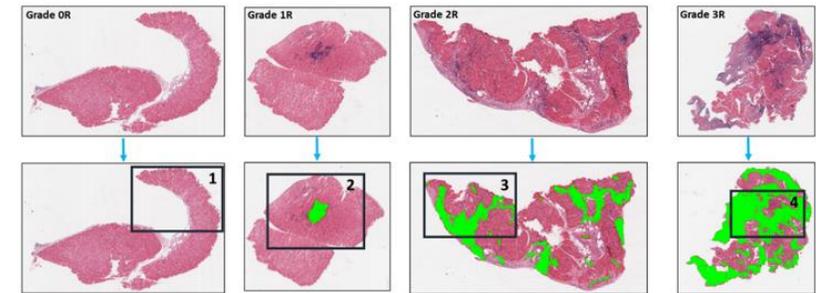
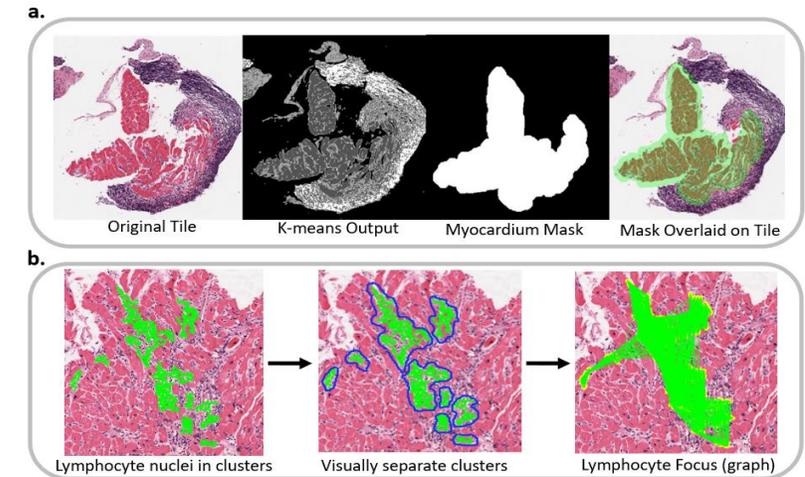
Especially grade group 2



- Doubling of cribriform had HR of 1.19 when controlling grade, stage, pre-operative PSA, age
- Model was similarly prognostic across four institutions
- C-index of 0.66 in grade group 2 patients with cribriform, potential role in active surveillance

An Automated Computational Image Analysis Platform for Accurate and Reliable Histologic Grading of Cardiac Allograft Rejection

- Studies demonstrate the poor reliability of ISHLT grading (kappa = 0.39)
- Overall Inter-pathologist agreement of 65-70%
- Inter-pathologist agreement of 28.4% at the higher grades of rejection (2R and 3R)
- Potentially affect immunosuppressive therapy decisions
- Computer-assisted cardiac histology evaluation (CACHE)-Grader
- N=2472 endomyocardial biopsy slides, 3 sites
- Features associated with interactions between myocytes, lymphocytes (counts, areas, spatial relationships)
- CACHE - 65.9%
- Inter-Pathologist agreement – 60%

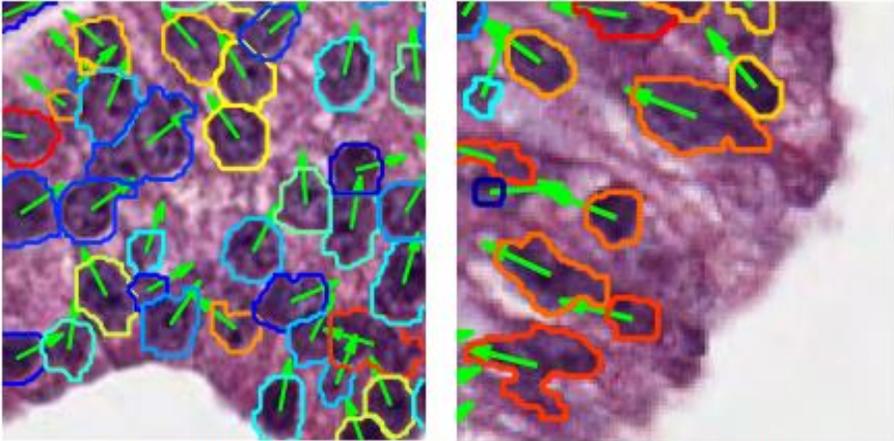
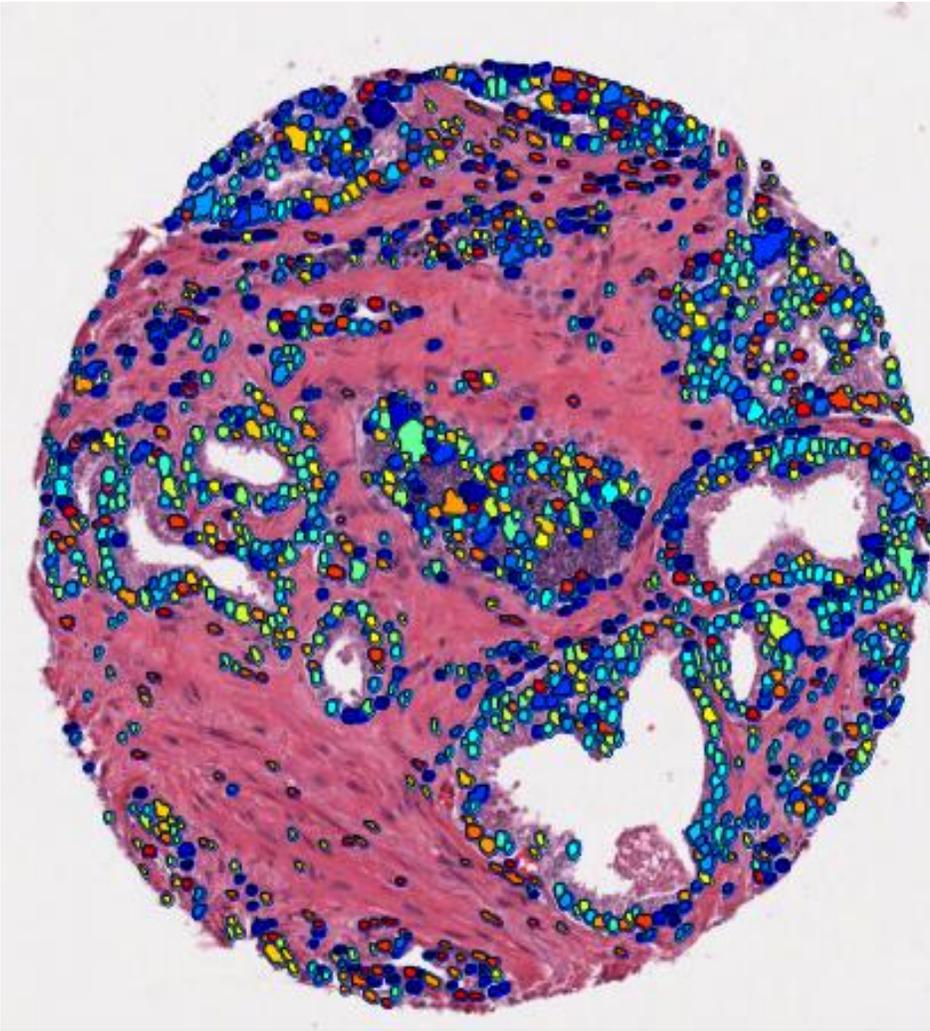


Combined Validation (S_1)

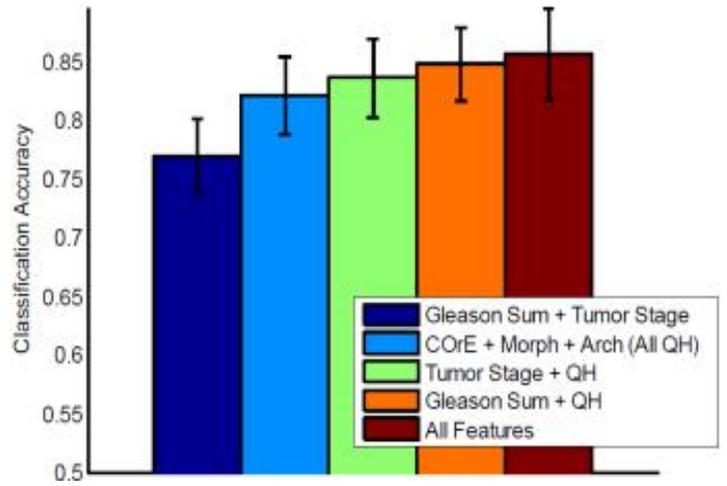
$S_1 + S_2 + S_3$

True Class \ Predicted Class	0R	1R	2R	3R
0R	420	183	20	
1R	28	405	183	17
2R		23	110	17
3R		2	21	18

Cell orientation entropy (COre) features stratify more and less aggressive prostate cancer on tissue microarrays

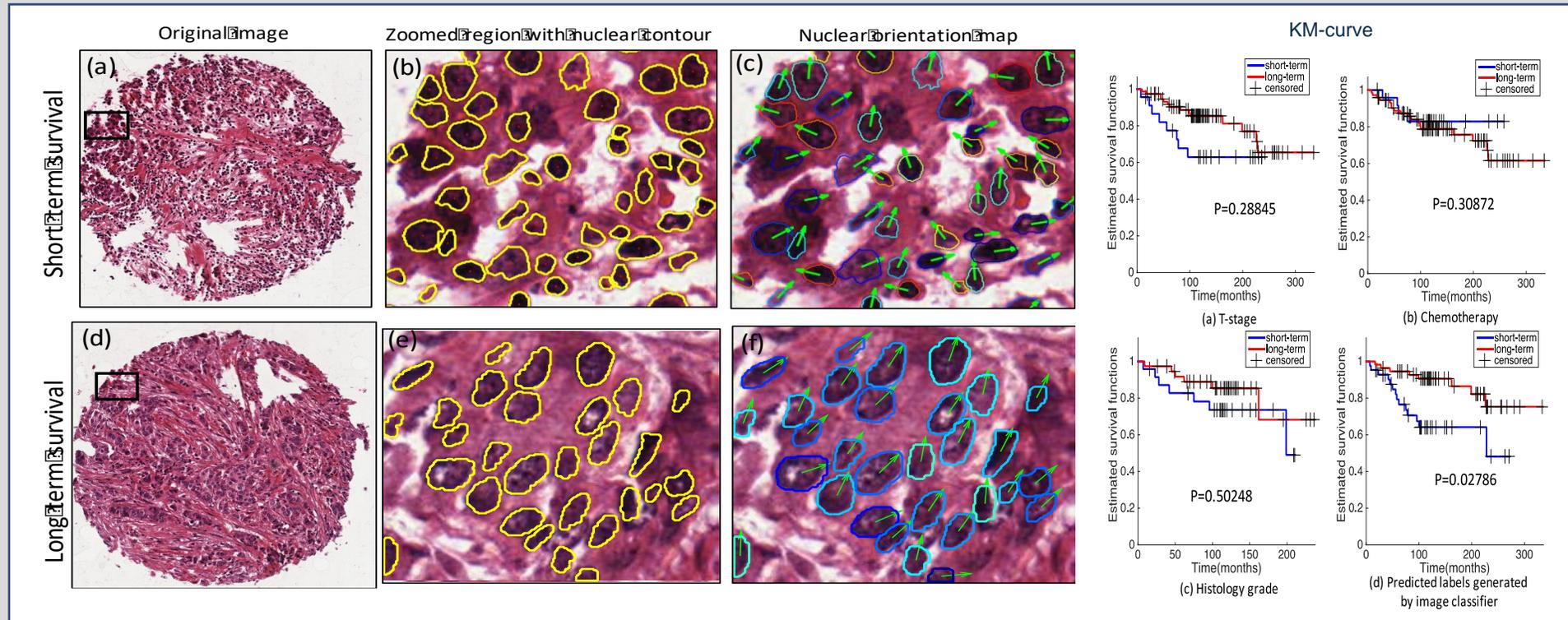


Aggressive cancer (left) shows more disorder in orientation of the nuclei compared to less aggressive cancer (right)



Lee, G, Ali, S, et al., "Cell Orientation Entropy (Core): Predicting Biochemical Recurrence from Prostate Cancer Tissue Microarrays", In Proc of Medical Image Computing and Computer Assisted Interventions (MICCAI), vol. 3, pp. 396-403, 2013.

Nuclear Shape and Orientation Features from H&E Images Predict Survival in Early Stage Estrogen Receptor Positive (ER+) Breast Cancers



Unmet Clinical Need

- Early stage ER+ breast cancer (BCa) is the most common type of breast cancer in the United States
- Identifying which patients will receive added benefit from adjuvant chemotherapy is important

Data

- TMA of 276 ER+ LN- patients
- Training cohort (n=177)
- Validation cohort (n=99)

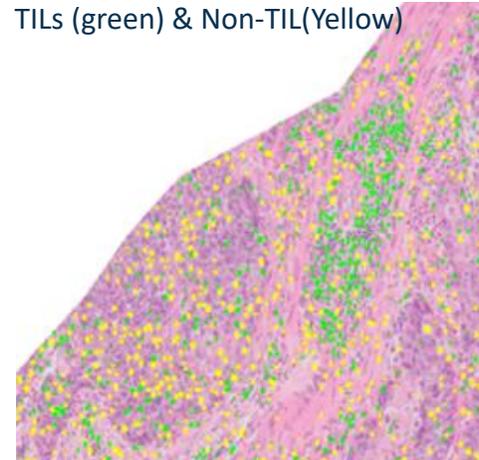
Spatial arrangement of tumor infiltrating lymphocytes (TILs) predict response to Nivolumab in non-small cell lung cancer (NSCLC)

Hypothesis: Spatial arrangement of TILs and local density variance are highly correlated to the patient response.

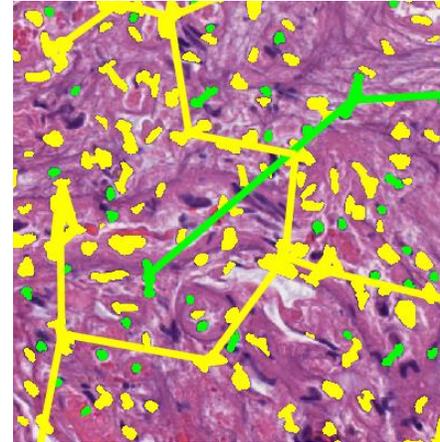
Data sets:

Two independent data (whole slide image) acquired from UPenn (32) and CCF (24)

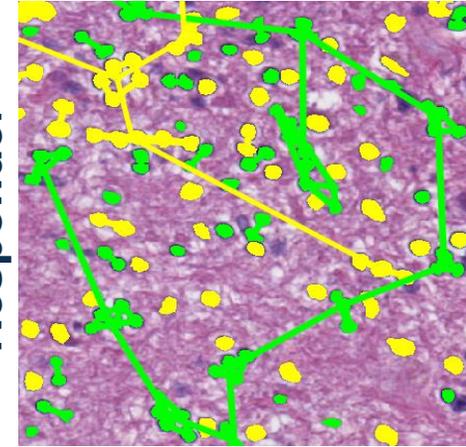
TIL detection and image feature extraction



Non-Responder

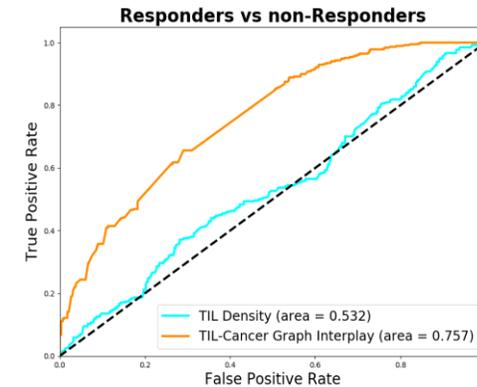


Responder



Top 5 most significant features obtained by feature selection

1. Median of TILs formed areas
2. Ratio of Cancer cells to TILs cells
3. Cancer cell averaged Density
4. Density of TILs
5. Median of Cancer cell formed areas



A QDA classifier was trained using a Training set (n=32) and a independently validation set from a different institution (n=24).

3. HistoSuite Tool Development

U01 NIH-NCI-ITCR



CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS

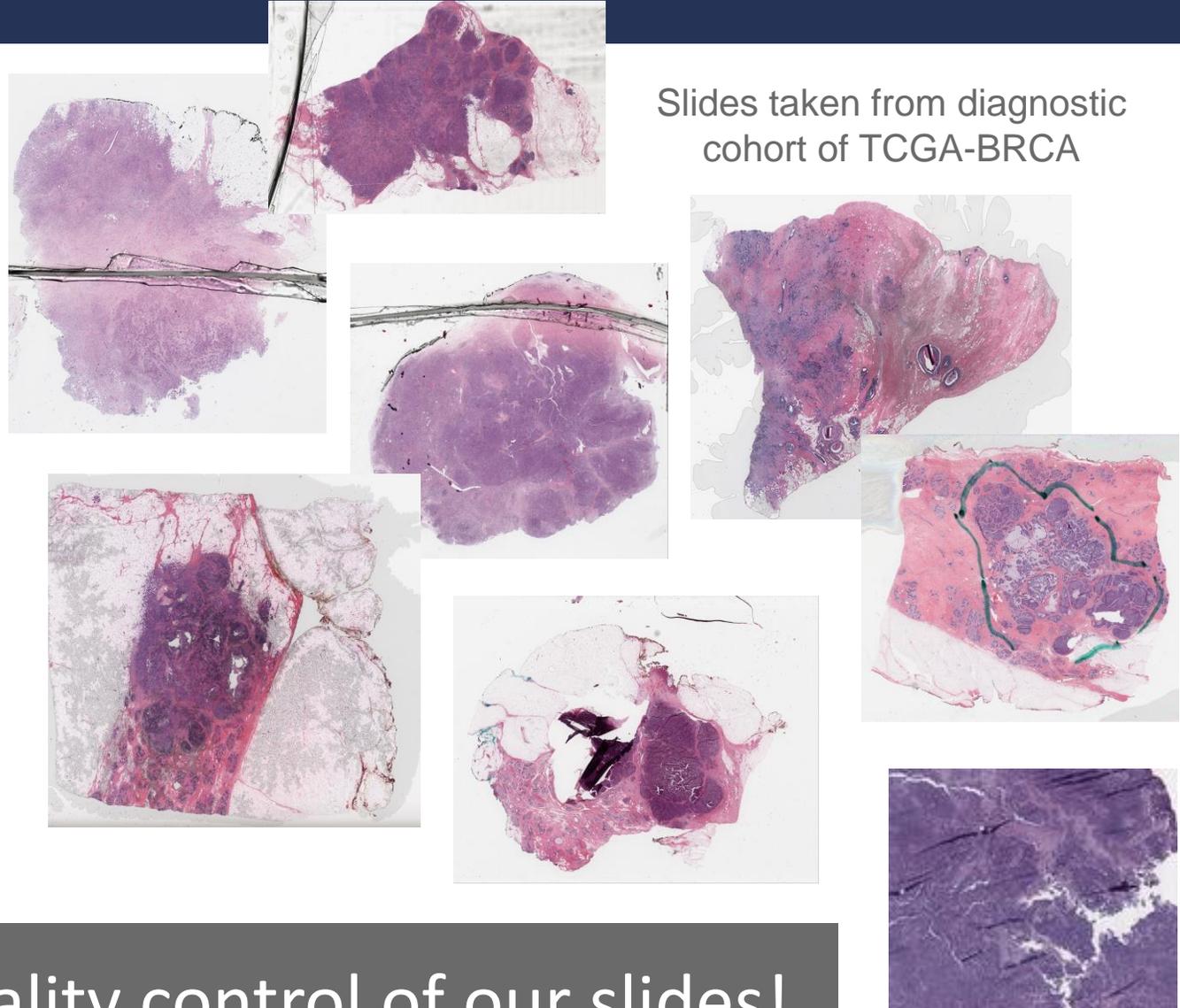
Unmet Need For Quality Control

- Transition to digital pathology workflows

- Digital Quality Control is paramount
- Recut and rescan slides immediately before getting into a workflow to a pathologist
- Cost and efficiency savings

- Previously not insurmountable

- Increasingly too time consuming to do manually
- Non-reproducible



Slides taken from diagnostic cohort of TCGA-BRCA

We need better quality control of our slides!

Surprising lack of reproducibility in manual QC

- For n=330 slides we simply provided a protocol and asked 3 readers:
- “Is this a good enough quality slide to computationally analyze?”
- We looked at the concordance between 3 readers
- This implies that each of these 3 readers would have started with a different dataset before even beginning their experiment
- Irreproducible QC = Irreproducible Experiments!

Stain	Agreement	Kappa	Gwet's AC1
Without HistoQC			
H&E	0.73	0.26	0.59
PAS	0.73	0.31	0.56
SIL	0.75	0.50	0.52
TRI	0.69	0.36	0.43
With HistoQC			
H&E	0.96	0.91	0.92
PAS	0.89	0.75	0.79
SIL	0.96	0.93	0.93
TRI	0.90	0.77	0.81

What is HistoQC?

- Open source reproducible slide quality metrics with artifact localization
- Python backend
 - identify artifacts and produce binary masks of “good” tissue
 - compute actionable quality scores and metrics
- HTML5 front end for visualizing and investigating results
- Able to aid in detection of Batch Effects!
- Available: <http://HistoQC.com>

The image shows a screenshot of a web browser displaying the HistoQC interface. At the top, there is a code editor showing a Python configuration file with the following content:

```
[pipeline]
steps= BasicModule.getBasicStats
      BasicModule.getMag
      ClassificationModule.byExampleWithFeatures:pen_markings
      #ClassificationModule.byExampleWithFeatures:pen_markings_red
      ClassificationModule.byExampleWithFeatures:coverslip_edge
      #LightDarkModule.getIntensityThresholdPercent:bubble
      LightDarkModule.getIntensityThresholdPercent:tissue
      #BubbleRegionByRegion.pixelWise
      LightDarkModule.getIntensityThresholdPercent:darktissue
      MorphologyModule.removeSmallObjects
      MorphologyModule.fillSmallHoles
      BlurDetectionModule.identifyBlurryRegions
      BasicModule.finalProcessingSpur
      BasicModule.finalProcessingArea
      HistogramModule.compareToTemplates
      HistogramModule.getHistogram
      BrightContrastModule.getContrast
      BrightContrastModule.getBrightnessGray
      BrightContrastModule.getBrightnessByChannelinColorSpace:RGB
      BrightContrastModule.getBrightnessByChannelinColorSpace:YUV
      DeconvolutionModule.seperateStains
      SaveModule.saveFinalMask
      SaveModule.saveThumbnails
      BasicModule.finalComputations
```

Below the code editor, the web interface shows a table of slide data with columns for filename, comments, levels, height, width, mpp_x, mpp_y, Magnification, pen_markings, coverslip_edge, nonwhite, and dark. The table contains three rows of data for different slide files.

filename	comments	levels	height	width	mpp_x	mpp_y	Magnification	pen_markings	coverslip_edge	nonwhite	dark
TCGA-A1-A05D-01Z-00-DX1.D8178FA9-0951-4248-91D2-44C28C6E95.svg		4	80287	94075	0.2521	0.2521	40	0.020845157541693442	0.01244133364784734	0.2598432039216643	0
TCGA-A1-A05D-01Z-00-DX1.A8717148-F964-4F29-88E2-97287C640432.svg		4	84072	92155	0.2521	0.2521	40	0.00051993251581759248	0	0.411151913519012993	0
TCGA-A2-A04U-01Z-00-DX3.06D17357-46A8-4DC3-A22B-2F4E86E3F79.svg		4	86726	130424	0.2465	0.2465	40	0	0	0.2399908201731674	0

Below the table, there is a bar chart showing various quality metrics for each slide. The chart has multiple series representing different quality metrics, with the x-axis corresponding to the slide files and the y-axis representing the metric values.

At the bottom of the interface, there are thumbnail images of the slides, showing the original histology images and the corresponding binary masks generated by HistoQC.



1. Janowczyk A., Zuo R., Gilmore H., Feldman M., Madabhushi A., “HistoQC: An open-source quality control tool for digital pathology slides”, JCO Clinical Cancer Informatics, 2019
2. Chen, Zee, Smith et al, Assessment of a Computerized Quantitative Quality Control Tool for Kidney Whole Slide Image Biopsies, Journal of Pathology, 2021

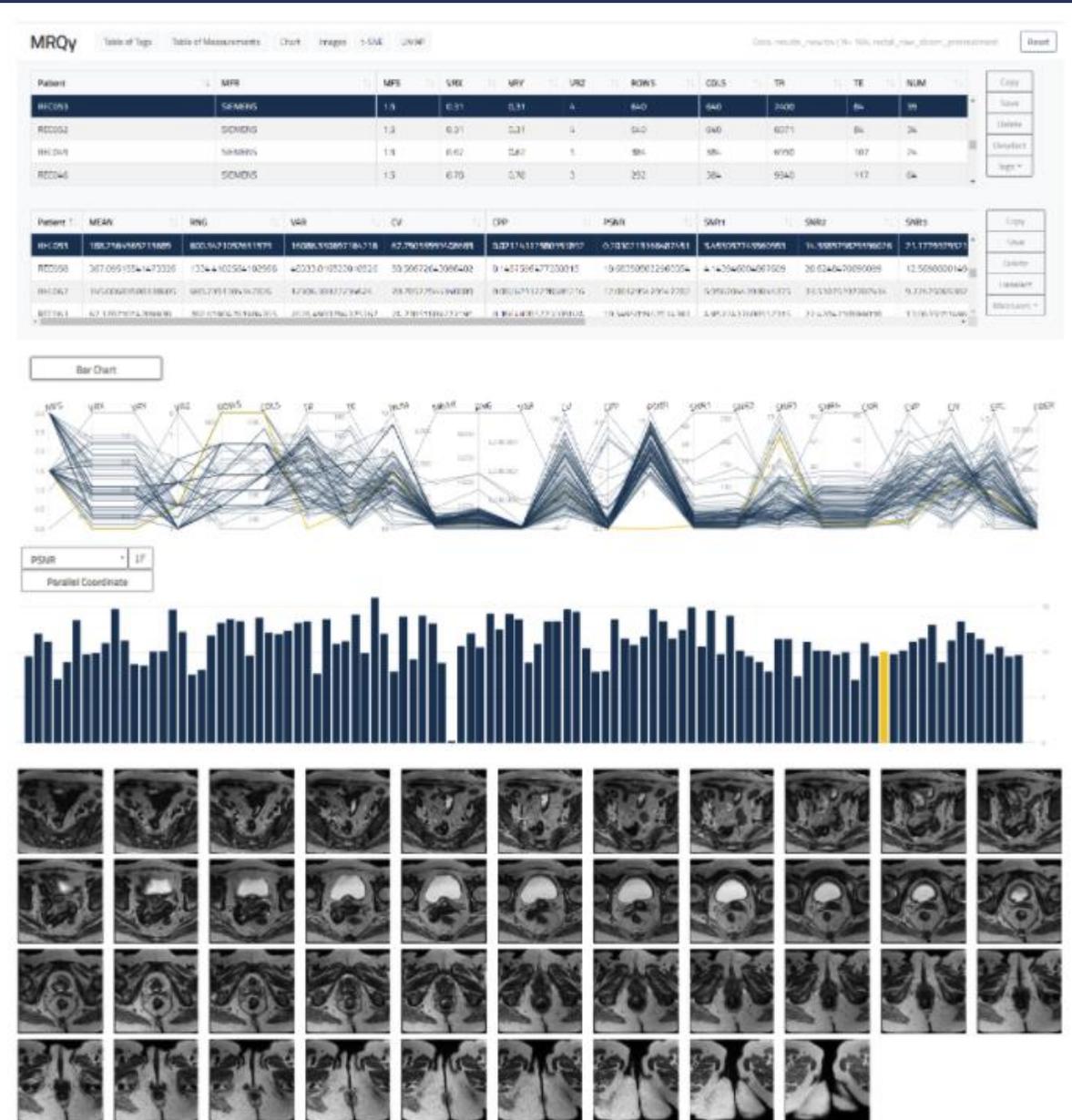
Extended HistoQC to imaging space with MRQy

Sadri A., Janowczyk A., Zhou R., Verma R., Beig N., Antunes J., Madabhushi A., Tiwari P. and Viswanath S., "Technical Note: MRQy -- An Open-Source Tool for Quality Control of MR Imaging Data", Medical Physics, 2020 (In press)

NIH ITCR U01 CA248226

RadXTools for assessing tumor treatment response on imaging

- <https://github.com/ccipd/MRQy>



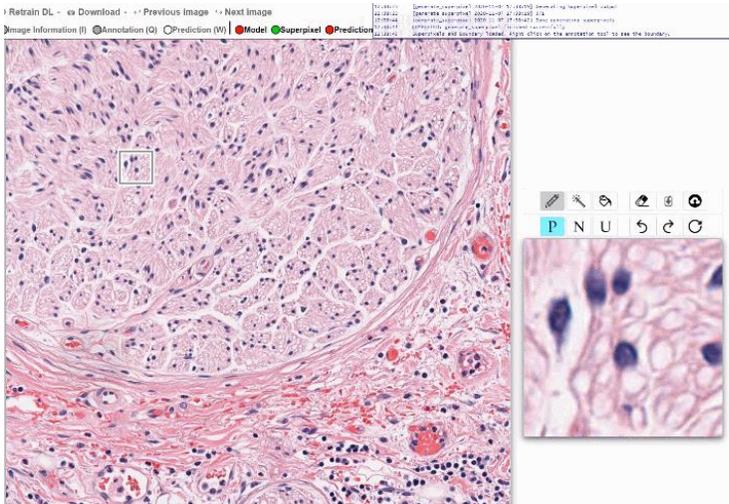
Quick Annotator

An open source digital pathology tool for rapidly annotating objects

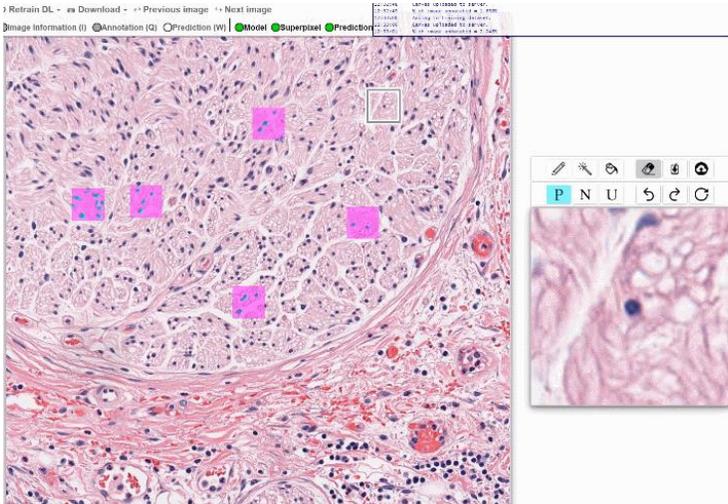


CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS

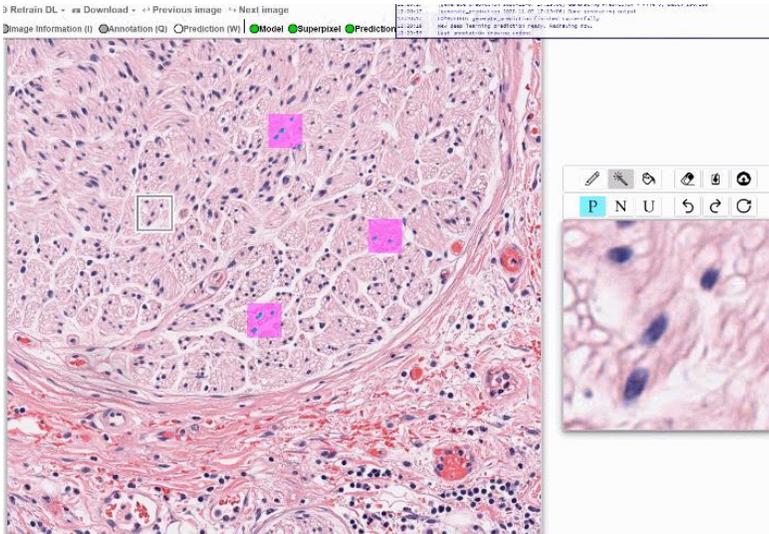
Quick Annotator Approach



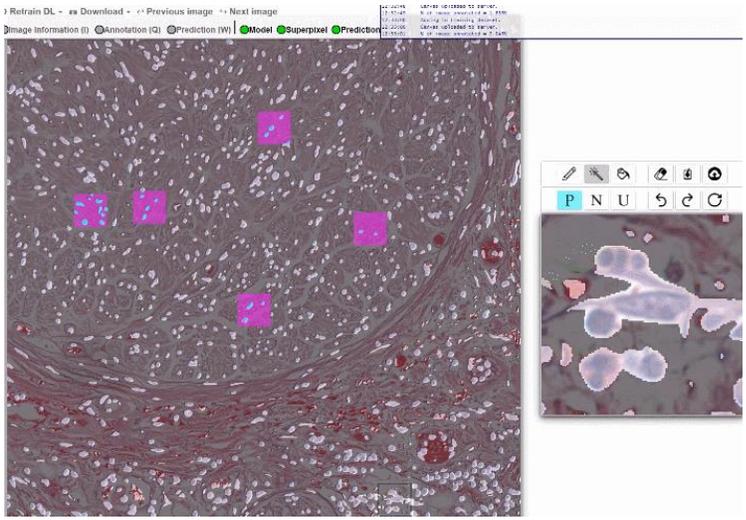
Annotating images



Deep learning prediction



Accept



Revision

Quick Annotator – Results

- Quick annotator significantly improves efficiency of annotation gathering
- Deep learning results in stain and domain agnostic tool
- Potential to improve upon human capabilities
- Tool Open Source for community usage and feedback
- Future work: support whole slide images

Tissue scale	Histologic structure	Number of slides	Number of ROIs	Number of histologic structures	QA total time	QA human time	Manual Time	Speed up	F-score
Small	Cell Nuclei	5	400	337,386	473	391	40,165	102X	0.97
Medium	Tubules	10	100	5,692	121	101	923	9X	0.95
Large	Epithelium	10	100	14,187	167	113	4,433	39X	0.89



https://www.youtube.com/watch?v=J34_ISZn-CM
<http://quickannotator.com>

Miao R., Toth R., Zhou Y., Madabhushi A., Janowczyk A. "Quick Annotator: an open-source digital pathology based rapid image annotation tool", The Journal of Pathology: Clinical Research, 2021

PatchSorter

A High Throughput Digital Pathology Tool for Cell Labeling



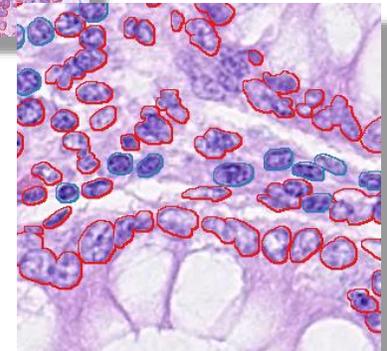
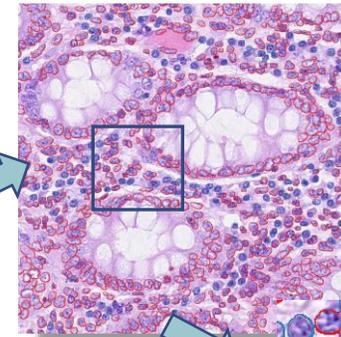
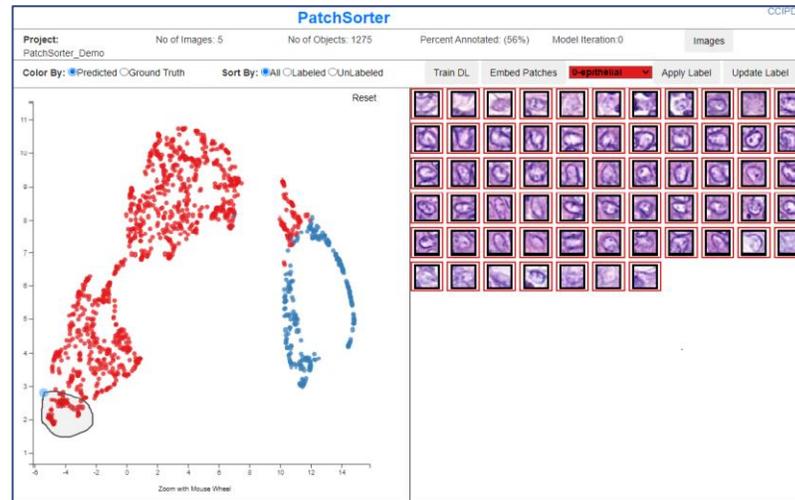
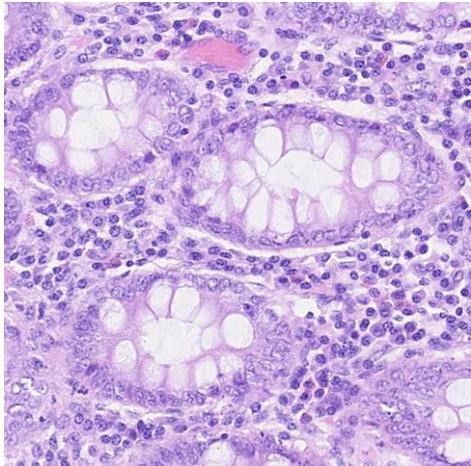
CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS

Motivation and Experimental Design

Motivation

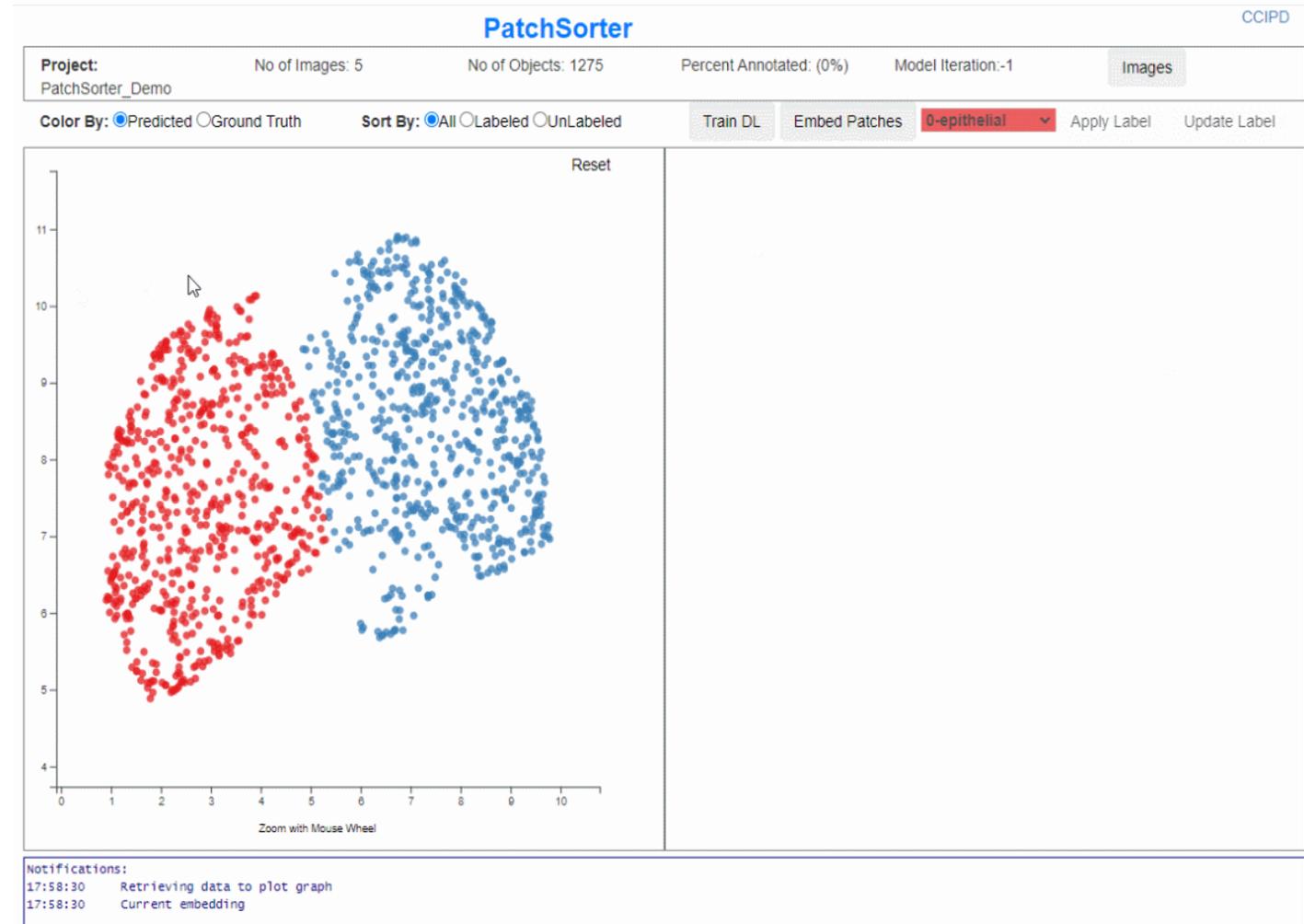
- Computational pathology often requires assigning class-types, or labels, to segmented cells.
- Manually labeling the millions of cells present in digital pathology images is intractable at scale.

- PatchSorter enables users to assign labels at a group, as opposed to individual cell level, greatly improving labeling efficiency by over 60%.
- As the backend deep learning model is trained, clusters become more distinct further improving efficiency.



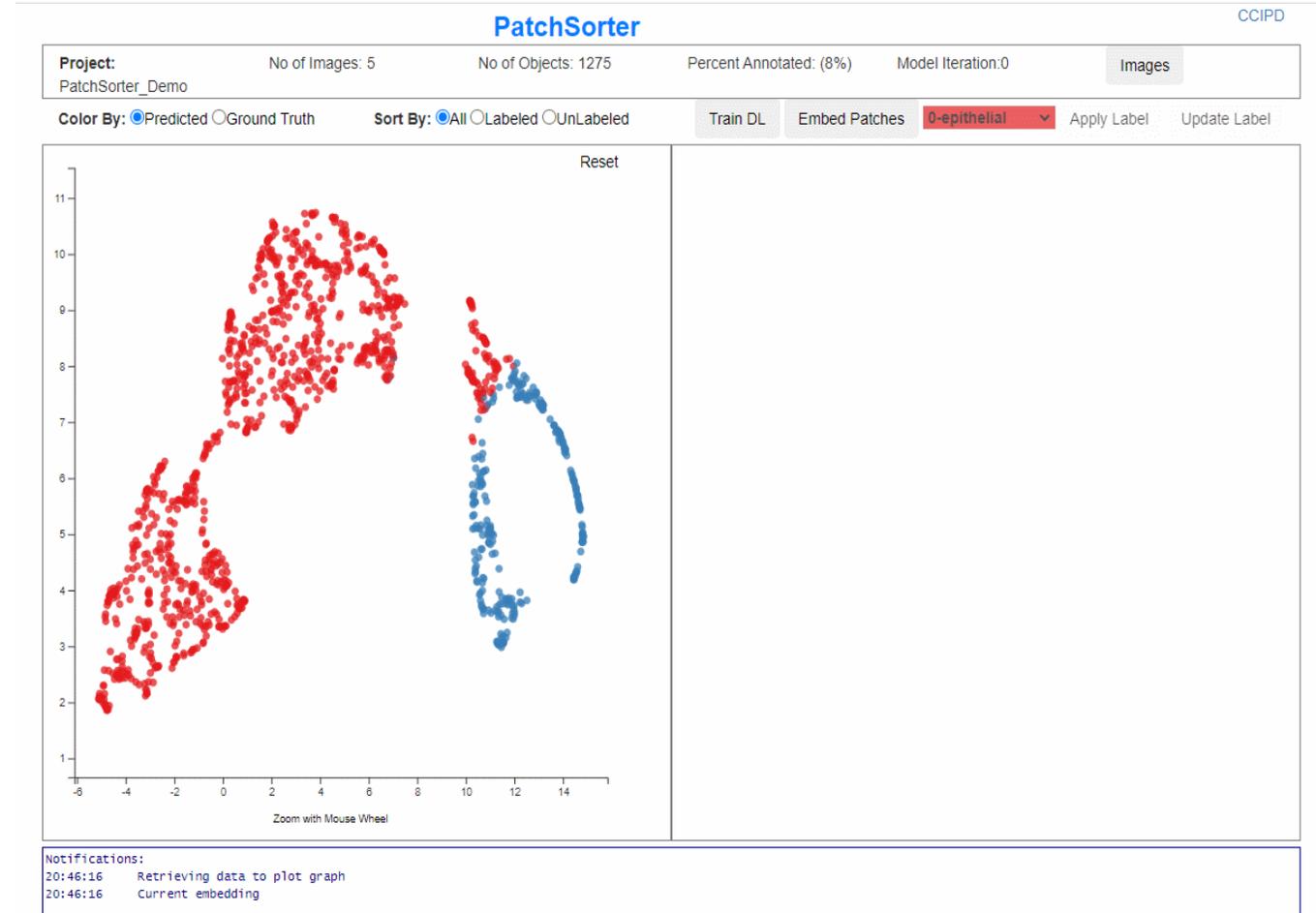
Collecting First Set of Labels

- A deep learning model is iteratively trained using provided labels to improve class separation.
- In the left plot, increased separation further facilitates rapid group selection and labeling.
- Options to view labeled or remaining unlabeled cells helps focus effort where needed.



After Deep Learning Model Training

- After importing images, an unsupervised embedding of cells into 2 predicted classes is visible in the plot (left).
- The user lassos points of interest in the plot and subsequently applies a definitive epithelial label (right, red-boxes).
- Importantly, similar cells appear near each other, enabling bulk selection, review, and labeling.



371% efficiency improvement for assigning labels to appropriate objects

Thank you!

Interested in
Collaborating?
Email me!

Email: andrew.janowczyk@case.edu

Digital pathology blog: andrewjanowczyk.com

HistoQC: <http://histoqc.com>

Quick Annotator: <http://quickannotator.com>

MRQy: <https://github.com/ccipd/MRQy>



CENTER FOR
COMPUTATIONAL IMAGING
AND PERSONALIZED DIAGNOSTICS



**Swiss Consortium
for Digital Pathology**